# SYSTEMATICS AND MOLECULAR PHYLOGENY OF MARINE PRAWNS AND SHRIMPS OF GUJARAT

A Thesis submitted to The Maharaja Sayajirao University of Baroda

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**Doctor of Philosophy** 

In

Zoology

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# List of abbreviations used throughout the text

М	:	Meter
Km	:	Kilometer
Mm	:	Millimeter
v/v	:	Volume/Volume
e.g.,	:	For example
i.e.	:	That is
viz.	:	that is to say; namely
GoK	:	Gulf of Kachchh
Gokh	:	Gulf of Khambhat
WoRMS	:	Word Register of Marine Sciences
Coll.	:	Collector
Ovi. female	:	Ovigerous female
Р	:	pereiopod (P1 to P5: First to fifth pereiopod)
Plp	:	Pleopod (P1 to P5: First to fifth pleopods)
Mxp1& Mxp2	2 :	First and Second maxilliped
Mxp3	:	Third maxilliped
CMFRI	:	Central Marine Fisheries Research Institute
DNA	:	Deoxyribonucleic acid
RNA	:	Ribonucleic acid

Mt DNA:Mitochondrial DNACOI gene:Mitochondrial cytochrome oxidase subunit 116S rRNA:16S ribosomal RNANCBI:National Center for Biotechnology InformationBoLD:Barcode of Life Datasystem

# **SUMMURY**

India is one of the 12 megadiverse countries of the world and supports 4 terrestrial biodiversity hotspots, highly biodiversity-rich, but endangered eco-regions (Myers et al., 2000). The coastline of India is about 7516.6 km long, with the mainland contributing 5422.6 km and the offshore islands contributing 2094 km (Andaman and Nicobar Islands: 1962 km; Lakshadweep Islands: 132 km) (Ahmad 1972; Kumar et al., 2006). The coastal area is divided into the west coast and the east coast. Both the coasts are significantly different in their geo-morphology. The Western coast of India is dominated by rocky shore habitat, while the east coast of India mostly has sandy beaches, mudflats, lagoons, and marshes. Crustaceans exist to the fourth-largest diversity, and they are the second most abundant diverse animal group on the planet. They abundantly inhabit the coastal marine environment. Gujarat is the western proximity of India and harbors the longest coastline of approximately 1650 km. The state's coastline is divided primarily into three coastal areas, *i.e.*, the Gulf of Kachchh, Saurashtra coast, and the Gulf of Khambhat. The coastline of Gujarat encompasses almost all types of intertidal habitat, from hypersaline estuaries, salt marsh, mudflats to sandy and rocky shores with every degree of exposure, and widely different profile. The subtidal habitats are equally diverse and rich.

Prawns and shrimps belong to Order Decapoda, infraorders Dendrobranchiata and Pleocyemata, respectively, and they are one of the most diverse and important groups of crustaceans. Prawns are the most significant food source with great economic importance as both capture and culture fisheries. Shrimps also have been attractive due to their great diversity throughout their evolutionary history and ornamental values. Some species may not have commercial value but are important to form an integral part of the food web of the tropical marine system. The commercial marine species are generally found in shallow or moderately deep-water regions along the continental shelves at less than 100m depth, and some are found even at nearly 5700 m depth. Many shrimp species are pelagic, but most of the species are benthic, living on a variety of hard and soft substrates like rock, mud, sand, shell particles, or a mixture of these fragments, and some species are symbiotically associated with others marine organisms.

Prawns and shrimps are a highly important group of marine decapods, and so far, 4048 species belonging to 471 genera are reported globally (Grave and Fransen, 2011). In India, so far, 364 species belonging to 128 genera are reported (Samuel et al., 2016). The maximum number of species are reported from the East coast of India compared to the West coast. As compared to the other coastal states of India's western coast, the prawn and shrimp fauna of Gujarat is less studied in terms of the intertidal species. In 2015, Trivedi et al. reviewed the literature available on the crustacean fauna of Gujarat and compiled a checklist. They have reported 30 species of prawns and shrimps belonging to 12 genera. It is noteworthy that all the earlier studies on the prawn fauna of Gujarat were focused on the commercial species (e.g., population study, stock assessment) as compared to the intertidal species. The landing centers of the Gulf of Kachchh and Saurashtra region are maximally explored compared to the other area. Only a few studies are carried out on the systematics of this group.

The present study was initiated with the following objectives.

- **1.** Study the Diversity, morphological taxonomy and distribution of marine prawns and shrimps.
  - I. Systematics of marine prawns and shrimps: A taxonomical approach
  - II. Distribution pattern and habitat preferences of prawns and shrimps in different coastal regions of Gujarat
- **2.** Establish the phylogenetic relationship among prawn and shrimp species.
  - I. Morphological phylogeny of prawns and shrimps: A cladistic analysis.

II. A comprehensive phylogenetic analysis of prawn and shrimp based on mitochondrial COI sequence data.

#### **Research Methodology:**

#### Study Area-Gujarat

The current research work has been conducted in the coastal region of Gujarat province. It is situated on India's western coastline bounded by the vast Arabian Sea on its three sides. Gujarat state is located between 20°06' N to 24°42' N latitude and 68°10' E to 74°28' E longitude, shares a North-Western border with Pakistan and Rajasthan, a North-Eastern border with Madhya Pradesh, and a South-Eastern border with Maharashtra. It has a 1,96,024 Km<sup>2</sup> total surface area (including Daman and Diu UT) and the longest coastline (about 1,650 km) of India. That accounts for 22% of the total coastline available to the country, with 164,200 km<sup>2</sup> of the continental shelf (35.3% of the country) and 214,000 km<sup>2</sup> of Exclusive Economic Zone (EEZ) (9.9% of the country).

Gujarat's total coastline consists of different marine habitats, including 29% of muddy flats followed by 28% sandy beach, 22% marshy coast, and 21% rocky coast. Gujarat's coastal area extends from the Western Ghats in Valsad (Umargam) to Kori creek (Kachchh) on the North-Western coast. The Gujarat state's coastal area is very different in terms of geomorphology from the rest of the West coast of India. The coastal zone of Gujarat state is divided into three major geographical parts, two major gulfs; namely Gulf of Kachchh and Gulf of Khambhat, and the Saurashtra coastline (fig. 1), each of which has its distinctive character, climate variation, and diverse geo-environmental features, which embrace diverse coastal habitats as well as ecological significant biota. The three regions vary a lot regarding tidal variation and marine habitat diversity.

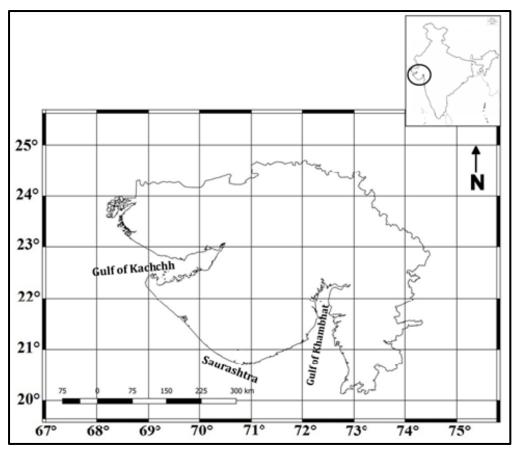


Figure 1 Map showing the division of study area into three major regions.

#### Selection of study sites

On the coast of Gujarat, a comprehensive field survey was conducted. The coastal region is divided into three major regions, and each coastal region is split further into the coastal districts. A total of 15 coastal districts were surveyed, and based on the survey, 70 sites were selected from three coastal regions of Gujarat for the samplings. However, due to repeated seasonal accessibility, permissions from Government authorities in Marine National Park and Defense Services Authorities in Kachchh district, 59 sites were finally selected for regular studies. Therefore, some of the critical island sites of Marine National Park and North Western creek habitats of Kachchh could not be surveyed. During the present study, the selections of the study sites from all the three major coastal regions have been based on the three parameters: one was habitat type (e.g., sandy shore, rocky shore, muddy shore, mangrove mudflats, and coral reef); the second was the

accessibility of the area, and the third one was the sample accessibility or availability.

#### Gulf of Kachchh

The Gulf of Kachchh is the region of peninsular Saurashtra-Kachchh in the western state of Gujarat and located between 22°15' N and 23°40' N Latitudes and 68°20' E and 70°40' E Longitudes. It is approximately 75 km wide, and 125 km long, narrow mouth funnel-shaped, East-West oriented indentation of Gujarat coast. On the northern sides, the Gulf starts with Kachchh point and lies between the beaches of Devbhumi Dwarka and Jamnagar districts in Saurashtra regions. The tides in the Gulf of Kachchh are mixed, predominantly semi-diurnal types with massive diurnal inequality. On the northern sides of the Gulf, the tides vary from 3 to 8 m, and on the southern sides, it's 3-5 m. The shape and orientation of the Gulf is the main reason for tide amplification. The high tidal range and tidal currents are important geological agents that play a significant role in sedimentation and shaping the land of the Gulf. The gulf comprises 4 coastal districts, namely, Kachchh, Rajkot, Jamnagar, and Devbhumi Dwarka. The narrow mouth funnel-shaped Gulf is covering an area of 7350 km<sup>2</sup>, has channel depths varying from 20 m at the head to 60 m at the outer Gulf. The average depth is approximately 30 m, and the minimum is up to 5 m (Gupta and Deshmukh, 2000). It is a semi-enclosed coastal area with diverse coral reefs and rocky ecosystem. The geophysical parameters suggest that the Gulf of Kachchh is quite different in many aspects than the Gulf of Khambhat. There are many major and minor islands in the Gulf of Kachchh; they support rich habitat diversity and biodiversity. According to official records, 42 islands in the GoK cover a total area of about 410.6 km<sup>2</sup>.

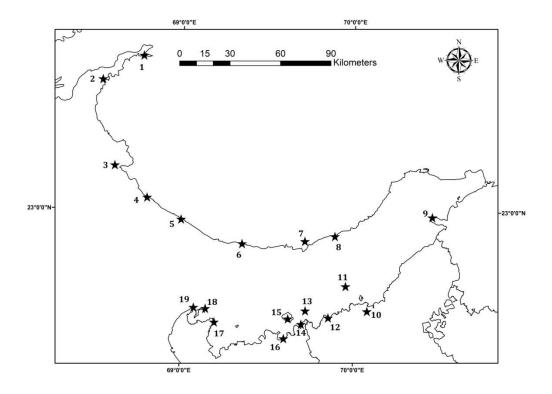


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A total of 19 sites from 4 coastal districts (Kachchh, Rajkot, Jamnagar, and Devbhumi Dwarka) situated along the Gulf of Kachchh were surveyed from the collection of samples (fig. 1.1). Vast patches of mangroves also exist along the northern shore of the Gulf. In the Gulf, the intertidal zone is rockysandy supra-tidal zone, muddy-rocky middle intertidal zone with boulders, and lower zone with diverse coral reef zone. The Kori creek to Jakhau and Mundra to Kandla Port are irregular and exclusively muddy with mangroves patches. Navlakhi to Jodiya has mudflats, while from Jamnagar to Okha (trending E-W), the coast has rocky shoreline (like rock mounds), the subtidal zone with water channels, submerged islands, sand bars, coral reefs, and mangroves. The GoK, Mithapur, Sivrajpur, and Dwarka are the only Gujarat areas where coral reefs exist. The GoK provides various kinds of habitats like coral reefs, mangroves, creeks, open mudflats (hard mud, soft mud and soup mud), islands, rocky shore, sandy shore, etc., which provide a suitable environment for a large range of fauna and flora.

#### Saurashtra Coast

The Saurashtra Coast is situated in the South-Western part of Gujarat and located between 20°72' N and 22°42' N Latitudes and 68°99' E and 70°98' E Longitudes. It is approximately 865 km<sup>2</sup> long, East-West oriented indentation of the Gujarat coast. It lies between the Gulf of Kutch on the north-west side and the GoKh on the south-east side and surrounded by the Arabian Sea on the south and south-west. The Saurashtra coastline is very distinctive from the Gulf of Kachchh and the Gulf of Khambhat. The high tide surges maximum up to 5 m at the Saurashtra coast. The Saurashtra peninsular comprises 6 coastal districts, namely, Devbhumi Dwarka, Porbandar, Junagadh, Gir-Somnath, and Amreli. The minimum depth is about 3 m, and the average depth is up to 5 m. The topography of the coast changes all along the Saurashtra coast. The majority of the coastline of the Saurashtra coast is rocky and sandy with randomly distributed patches of muddy regions with heavy sediment load.

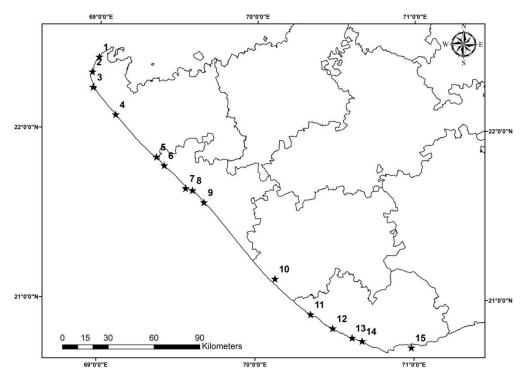


Figure 1.2 Map showing the sampling sites along the coastline of the

Saurashtra coast; 1-Okha;2-Mithapur; 3-Shivrajpur4-Okhamadh; 5-Harshad; 6-Miyani; 7-Kuchhadi Reef; 8-Porbandar; 9-Odadar; 10- Mangrol; 11-Veraval; 12-Sutrapada; 13-Dhamlej; 14-Kodinar; 15-Diu.

A total of 15 sites from 4 coastal districts (Devbhumi Dwarka, Porbandar, Junagadh, and Gir-Somnath) were surveyed for the collection of samples (fig.1.2). The intertidal zone of the Saurashtra coastline is unique in that the upper intertidal zone is rocky, somewhere sandy with mud, the middle intertidal zone is mostly rocky, with crevices, and the lower intertidal zone is mainly rocky with sparse sand. In the mid intertidal zone, tide pools are commonly found. The south coast of Saurashtra from Dwarka- Kodinar is about 250 km patch, with straight and smooth sandy or rocky-sandy beaches. The Saurashtra coastline is rocky-sandy, the east and west (E-W) with Continuous beach with projecting rock mounds, wave-cut platforms, occasional cliffs, the central part is sandy, and more rocky-muddy beaches are found in the eastern part. The formation of milliolite limestone along the coast is remarkable. Due to its unique geomorphologic properties and hydrodynamic processes, it provides a suitable environment for a wide range of marine fauna and flora. The intertidal zone of Saurashtra coasts is not very wide; generally, dynamic wave action is seen due to this reason.

#### Gulf of Khambhat

The Gulf of Khambhat is situated between the Saurashtra peninsula and the mainland of Gujarat state and located between 20°30' N and 22°20' N Latitudes and 71°45' E and 72°53' E Longitudes. It is approximately 131 km long and 70 km wide, wide inverted mouth, narrow end funnel-shaped North-South oriented indentation of Gujarat coast. On the western side, the Gulf begins from Amreli Point and lies between Surat and Valsad districts on the eastern side. The Gulf is best known for its extreme tides (maximum 12 m), which vary significantly in height and run into excellent speed 3 m/sec. On the western sides of the Gulf, the tides vary from 3 to 12 m, and on the eastern sides, it is 6-10 m, and the shape of the Gulf is the main reason for tide amplification. The Gulf of Khambhat comprises 8 coastal

districts Amreli, Bhavnagar, Ahmadabad, Anand, Bharuch, Surat, Navsari, and Valsad. The funnel-shaped Gulf, which covers an area of 3,120 km<sup>2</sup> comprised mainly of mudflats with some rocky area, is shallow with depths varying from 5 m at the head to 40 m in the channels. The average depth is about 20 m, and the minimum is up to 5m (Gupta and Deshmukh, 2000). There are a few sandy patches that are also observed intermittently.

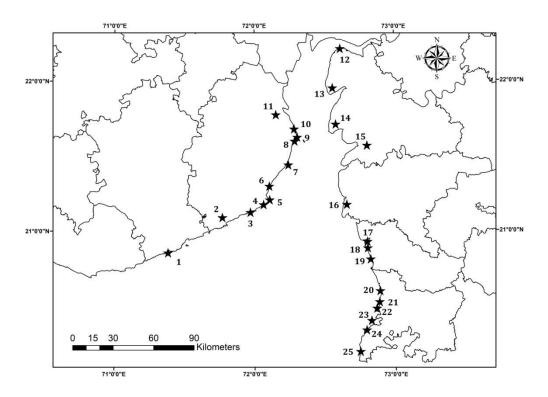


Figure 1.3 Map showing the sampling sites along the coastline of the Gulf of Khambhat; 1-Jafrabad; 2-Mahuva; 3-Unchakotda; 4-Jhanjhmer; 5-Sartanpar; 6-7-Gopnath; 8-Alang; 9-Koliyak; 10-Kuda; 11-Gogha; 12-Bhavnagar; 13-Kamboi; 14-Nada; 15-Dahej; 16-Hansot; 17-Hazira; 18-Vansi-Borsi; 19-Dandi; 20-Onjal; 21-Thithal; 22-Umersadi; 23-Udwada; 24-Daman; 25-Fansa; 26-Umargam.

A total of 26 sites from 6 coastal districts (Amreli, Bhavnagar, Bharuch, Surat, Navsari, and Valsad) situated along the Gulf of Khambhat have surveyed for the collection of samples (fig. 1.3). The rocky beaches (some rocky patches) are commonly found from Mahuva to Gopnath, reducing towards Ghogha and Bhavnagar. The Gulf receives freshwater by several inlets like major rivers, especially in monsoons such as Sabarmati, Mahi, Narmada, Tapi, Shetrunji, and many minor rivers. All the rivers from estuaries and their inflow carry solid suspended sediments into the Gulf. A medium-sized delta is present near Sartanpur, known as the Shetrunji River delta between Gopnath and Ghogha. The excessive turbulence churns the seabed and produces enormous quantities of silt and clay, making the seawater turbid and brownish, so that the light rays are impermeable. The Gulf's marine ecosystems, comprising mangroves, estuaries, creeks, and vast intertidal mudflats, are known to have rich biodiversity and several endemic flora and fauna.

#### Sample collection

The sampling was done from 59 sites along the Gujarat coastal area from 2015 to 2019 (figs. 1.1, 1.2, & 1.3). Various methods were employed for the collection of prawn and shrimps. Hand-picking and the handheld net method were adopted to collect intertidal shrimp species during the low tides. The fish landing center and local fish markets were also visited for the collections, where the trawler catch was examined for commercial species of prawn. The fish landing centers location, type of fishing trawlers, approximate depth of sample collected, gears used, etc. all this information was collected from the fishermen. The fresh specimen's photographs were taken immediately after the collection for bright and fresh coloration, and then the samples were preserved and brought to the laboratory.

# **Relaxation of specimens**

All the live samples were narcotized first to avoid dissection and diagnosis difficulties during the morphological identification. Narcotization was obtained with menthol crystal and by chilling and freezing the live specimens. For a few minutes, the specimen was submerged in the solution. After this, the samples were ready for preservation or morphological analysis.

# Preservation for Morphological Analysis

Samples for the morphological examination were immersed in 4% formalin for 4-6 hours and later transferred in 10% formalin (v/v). Formalin denatures DNA by rendering it unavailable by binding the surrounding histone proteins so that the isolation of DNA for genetic research is more complicated and, in some cases, impossible.

#### Morphological Identification and Deposition

In order to identify each specimen up to the species level using morphological characters (Annexure 1), the following steps were considered. The coloration was observed immediately after the collection of samples. Detailed morphometry and sex determination were carried out for each specimen. The taxonomically important body parts were dissected and examined under a stereomicroscope. All the specimens were identified up to the species level using various identification keys, monograms, and taxonomic literature. The detailed sketches of body parts were prepared for different species, and photographic identification plates were prepared. The total length (TL- tip of the rostrum to tip of telson) and carapace length (CL-tip of the rostrum to the posterior end of the carapace) were measured for all the specimens of every species using the digital vernier caliper ( $\pm 0.1$ mm accuracy). Smaller samples, mostly caridean, were measured using the stereomicroscope equipped with the measuring tool. For the validation of taxonomic classification, the species detailed were compared with the details available on Marine Species Identification Portal Website (www.speciesidentification.org), and the classification was adopted from the world Registered of Marine Species website (www.marinespecies.org). All the identified species were deposited into the Museum at the Department of Zoology, Faculty of Science, The Maharaja Sayajirao University of Baroda, Gujarat, India, with a unique museum accession code (e.g., ZL-AR-PR-1: *Metapenaeus affinis*) assigned for each species. The clear label was given to the specimen bottles with species accession number, scientific name, family name, collection site, and collector's name. All the families, genera as well as the species, and their synonyms are listed

following the criteria of WoRMS, and the taxa within the families are ordered alphabetically.

Since the coast of Gujarat is very long and has distinct and widely variable habitats, the species distribution was recorded in two different ways:

1. Geographical region-wise distribution of the species in the Gulf of Kachchh, Saurashtra, and Gulf of Khambhat.

2. Habitat wise distribution to understand the ecological role of the shrimp species.

#### 1 Region-wise distribution

The coastline of Gujarat is divided into three major regions, namely the Gulf of Kachchh, the Saurashtra coast, and the Gulf of Kachchh. All three areas have distinctive characters and habitat variations. There is a total of 15 coastal districts of Gujarat, where the Kachchh has the longest coastline. Gulf of Kachchh is bound by 4 districts, Saurashtra coast by 6 districts, and the Gulf of Khambhat by 8 districts, including the lower south Gujarat extension. During the sample collection at each sampling site, the prawn and shrimp species present were listed with latitude and longitude information. All the data were maintained in the excel sheets for the analysis.

#### 2 Habitat-wise distribution

During 2015 to 2019 field surveys, the coastal regions of Gujarat were conducted to study the habitat preference and distribution of the species. In the present study, the habitats are divided mainly into types of the shore (macro-habitat) as rocky, sandy, mudflats, and sub-littoral or pelagic regions of the coastal ecosystem (fig. 2). At each sampling site, the prawn and shrimp species present were listed, dominant species of animal and type of shore were notated. The zonation pattern is the most important phenomenon observed in the intertidal area. Here in the present study, different zones have been identified based on the presence of the dominant

animal community. The microhabitat preference by the intertidal species was also studied. On the sighting of animals, micro-habitat type and zone type were recorded.

*Micro-habitat classification-* Macro-habitats (types of shore) were further divided into six different micro-habitats (figs. 3 & 4).

**EnZ** (**Endozoic**): Always recorded in an internal association with a particular species of animal.

**EZ (Epizoic):** Always recorded in an external association with a particular animal species (e.g., coral reef and sea anemone).

*H* (*Hard Substrate*): Recorded associated with hard substrates (e.g., rock).

M (Mangrove): Recorded amongst mangroves.

**OM (Open Mudflats):** Recorded in the areas of open mudflat coast.

*SS (Soft Substrate)*: Recorded associated with soft substrates, sand, or mud tide pools (e.g., sand and mud).



Figure 2 Macro-habitats or intertidal habitats: a) Rocky shoreline at Harshad (Saurashtra); b). Muddy shoreline at Kambhoi (Gokh); Sandy shoreline at Mandvi (GoK).

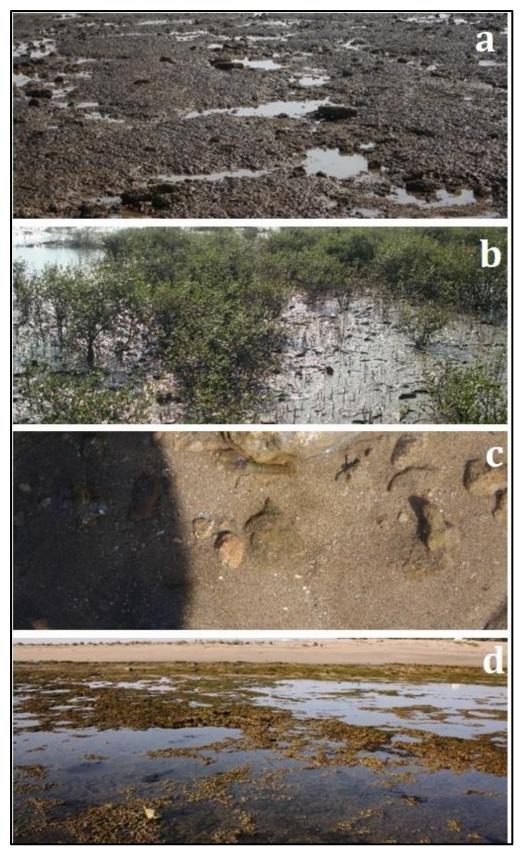


Figure 3 Type of micro-habitats or intertidal habitats: a) Open mudflat; b). Mangrove mudflat; c) Soft substrate; d) Hard substrate.

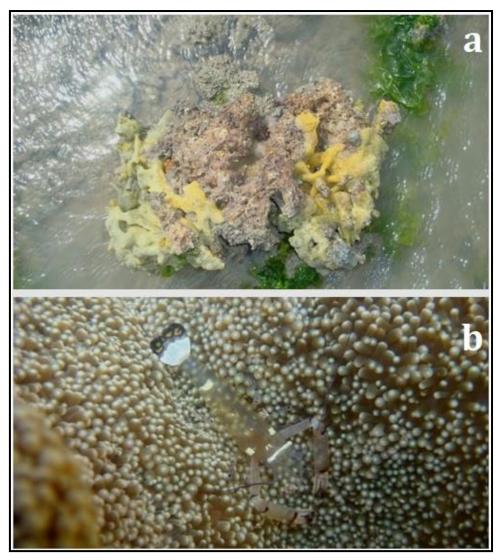


Figure 4 Types of micro-habitats or intertidal habitats: a) Endozoic; b) Epizoic.

# Specimen Information

The morphometric analysis includes 152 specimens belonging to 39 valid species (Table 1). For the morphometric analysis, 4 families viz. are Callianassidae Alpheidae, Upogebiidae, and Spongicolidae were omitted because the morphological structure is different. Family Pandaloidea was also not considered because, in this species, the only ovigerous females were reported during the present study. After identifying species belonging to suborder dendrobranchiate and pleocyemata shrimp, from each species of Penaeidae, three male samples were taken, and the female omitted due to the size variation. In the case of caridean shrimps (in few species), female specimens were also considered due to a smaller number of samples were reported.

# Multivariate Analysis of Morphometric characters

The morphometric measurements were taken based on the truss network system shows in fig 5 (Aktas et al., 2006). In this system total of eighteen landmarks and forty variables were measured. The total length (TL) was also calculated from the tip of the rostrum to the posterior end of the telson, and forty-one variables (Table 1) were measured using a digital vernier caliper (accuracy  $\pm 0.1$  mm). Small specimens were measured using a stereomicroscope equipped with a measuring tool. All the measurements were taken from thawed samples.

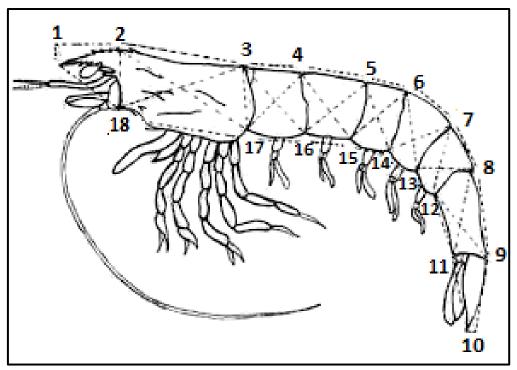


Figure 5 Truss network system used for morphometric analysis among prawn and shrimp species.

The average value was calculated. In this chapter, a significant correlation observed the average measurement of each variable and the total length. Therefore, it was necessary to remove the effect of size variation on species. To eliminate the variation resulting from each morphometric variable, they were standardized, according to Elliott et al. (1995).

$$Madj = M(Ls/Lo)^{b}$$

M is the original (primary) measurement; Madj is the size adjusted measurement of each variable. Ls is the overall mean of the standard length of all samples, and Lo is the standard length of the model or sample. Parameter b was estimated for each character from the observed data as the slope of the regression of log M on log Lo, using all prawn and shrimp group. The Ls/Lo values computed in the MS excel sheet and detecting the b parameter was carried out using the PAST (Paleontological Statistics Software) package version 3.25. The cluster analysis was performed UPGMA multivariate method to evaluate the morphometric relationship among all species in each order. In this chapter, hierarchical clustering analysis was represented as a dendrogram, where a joint of the tree illustrated each step in the clustering formation. The single linkage method was used with the Euclidean distance (statistic tool that quantifies the extent to which species within-cluster are similar) with 1000 bootstrapped.

Table 1 List of the morphometric variables used for the study of the
taxonomical relationship of the prawn and shrimp species

S. No.	No Variables*	Variable Name
1	var1-2	Rostrum length
2	var2-3	Carapace dorsal
3	var3-4	First Abdominal segment dorsal
4	var4-5	Second Abdominal segment dorsal
5	var5-6	Third Abdominal segment dorsal
6	var6-7	Fourth Abdominal segment dorsal
7	var7-8	Fifth Abdominal segment dorsal
8	var8-9	Sixth Abdominal segment dorsal
9	var9-10	Telson dorsal
10	var10-11	Telson ventral
11	var11-12	Sixth Abdominal segment ventral

12	vae12-13	Fifth Abdominal segment ventral	
13	var13-14	Fourth Abdominal segment ventral	
14	var14-15	Third Abdominal segment ventral	
15	var15-16	Second Abdominal segment ventral	
16	var16-17	First Abdominal segment ventral	
17	var17-18	Carapace ventral	
18	var1-18	Diagonal of anterior end carapace to	
		rostrum end	
19	var2-18	Perpendicular Carapace 1	
20	var2-17	Diagonal of Carapace 1	
21	var3-18	Diagonal of Carapace 2	
22	var3-17	Perpendicular Carapace 2	
23	var3-16	Diagonal of first Abdominal segment 1	
24	var4-17	Diagonal of first Abdominal segment 2	
25	var4-16	Perpendicular of first Abdominal segment	
26	var4-15	Diagonal of second Abdominal segment 1	
27	var5-16	Diagonal of second Abdominal segment 2	
28	var5-15	Perpendicular of second Abdominal	
		segment	
29	var5-14	Diagonal of third Abdominal segment 1	
30	var6-15	Diagonal of third Abdominal segment 2	
31	var6-14	Perpendicular of third Abdominal segment	
32	var6-13	Diagonal of fourth Abdominal segment 1	
33	var7-14	Diagonal of fourth Abdominal segment 2	
34	var7-13	Perpendicular of the fourth segment	
35	var7-12	Diagonal of fifth Abdominal segment 1	
36	var8-13	Diagonal of fifth Abdominal segment 2	
37	var8-12	Perpendicular fifth Abdominal segment	
38	var8-11	Diagonal of sixth Abdominal segment 1	
39	var9-12	Diagonal of sixth Abdominal segment 2	
40	var9-11	Perpendicular of fifth Abdominal segment	

#### Sampling of Molecular data

A total of 112 specimens of marine prawn and shrimps were collected for DNA examination. The samples represented 52 species of shrimps and prawns from different habitat and localities along the coastal area of Gujarat, including Union territory Daman and Diu.

#### DNA extraction

Genomic DNA was extracted from abdominal tissue or pleopods of prawns and shrimps. The initial weight was approximately 20 mg, and extraction was carried out using DNeasy Blood and Tissue kit (Qiagen) (Annexure 2).

#### Quantification of genomic DNA

After extraction of genomic DNA, quantification was carried out using QIAxpert (QIAGEN). Purity and concentration of DNA were measured using gel electrophoresis (Annexure 3).

# PCR amplification

The region of the mtCOI gene was amplified using primers given in (Table 2). The final volume of 20  $\mu$ l containing 10  $\mu$ l Taq PCR master mix (HiMedia), 10 pmol forward primer, 10 pmol reverse primer, 2  $\mu$ l template DNA and nuclease-free water (to make up the final volume). The gene amplification was carried out in a thermal cycler (Applied Biosystems Veriti®) using the PCR condition given in table 3 and 4.

Table 2 Oligonucleotide primers used in the present study to amplify the COI gene.

Primer	Primer sequence 5'-3'	Reference
LCO 1490	GGTCAACAAATCATAAAGATATTGG	Folmer et al., 1994
HCO 2198	TAAACTTCAGGGTGACCAAAAAATCA	

C1-J-1718	GGAGGATTTGGAAATTGATTAG	Simon et al., 1994	
C1-N-2191	CCCGGTAAAATTAAAATATAAACTTC		

Table 3 Thermal cycling conditions used in the present study to amplify the COI gene using LCO 1490 and HCO2198.

Stage	Stage 2		Stage 3		Stage 4			
1	(35 Cycles)		(35 Cycles) (5 Cycles)					
94°C	94°C	45°C	68°C	94°C	50°C	68°C	68°C	4°C
1	1min.	1	1 min.	1	1 min.	1 min.	10	$\infty$
min.		min.	30	min.	30	30	min.	
		30	sec.		sec.	sec.		
		sec.						

Table 4 Thermal cycling conditions used in the present study to amplify the COI gene using C1-J-1718 and C1-N-2191.

Stage 1	Stage 2			Stage 3	Stage 4
	(35 Cycles)			(1cycle)	
95°C	95°C	50°C	72°C	72 °C	4°C
3 min.	30 sec.	30 sec.	1 min. 30 sec.	7 min	$\infty$

# **Qualification and Quantification of PCR**

After PCR amplification, the amplified regions were observed using gel electrophoresis (Annexure 3).

# PCR Purification

Positive PCR amplified were purified, suing EXOSAP IT (affymetrix) according to the manufacturer's instructions. In a new PCR tube, 10  $\mu$ l of positive PCR product and 4  $\mu$ l of EXOSAP were added. A thermal cycler was run using PCR condition given in table 5.

Table 5 PCR purification condition used in the present study.

Stage I	Stage II	Stage III	
37°C for 15 minutes	80°C for 15 minutes	4°C	

## Sequencing

The PCR amplified product was bi-directionally sequenced using the same set of the universal primers (LCO 1490 HCO 2198 and COI 1718F COI 2191R) on the ABI 3730x196 capillary DNA analyzer using Big Dye Terminator v 3.1 sequencing kit at Eurofins, Bangalore.

## Sequence and Phylogenetic analysis

The qualities of the bi-directional chromatogram of the generated sequences were checked, and noisy parts were trimmed at both ends to avoid the noisy part using sequences nucleotide sequence DNA baser assembler 5.15 version. Further, each sequence was a check for sequence similarity through BLASTn analysis with Gene bank sequences (NCBI database). All the sequences were submitted on the NCBI portal with a unique accession code. The multiple sequences alignment was done for all the species of order Dendrobranchiata and Pleocyemata by multiple sequences alignment was opting using the MEGA X version. All missing data and gaps were eliminated from the data set. The phylogenetic was performed using the ML (Maximum Likelihood) method in the Kimura 2-parameter model with 1000 bootstrap replicates using the MEGA X version (Kumar et al., 2018).

#### **Key findings**

- The present study of marine prawns and shrimps' taxonomy and systematics revealed 52 species belonging to 27 genera and subgenera, 11 families, and 4 superfamilies.
- Among all the families recorded, the family Penaeidae and Alpheidae were dominant in the study area.
- Three species of shrimps of suborder Pleocyemata like *Gilvossius rotundicaudatus, Alpheus chiragricus,* and *Athanas parvus* were recorded for the first time from India.
- *Microprosthema validum* know as a rock lobster, was the first time reported from India's West Coast.

- Three species of caridean shrimp *viz. Synalpheus coutierei, Thor amboinensis,* and *Procletes levicarina* were reported first time from the western coast of India
- In the present study, species like *Megokris granulosus, Megokris sedili, Parapenaeus fissuroides indicus, Solenocera choprai, Latreutes anoplonyx, Alpheus lobidens, Alpheus malabaricus, Alpheus edwadrsii, Alpheus pacificus, Lysmata vittata, Palaemon pacificus, Palaemon serrifer,* and *Cuapetes grandis* were first time reported from Gujarat.
- So, the present study adds three more species to the list of India's prawns and shrimps, 4 more species to the checklist of India's western coast, and 22 more species to the checklist of prawns and shrimp of Gujarat.
- The study is divided into three major regions based on the distinctive character and habitat variation.
- The maximum diversity was observed from the Saurashtra coast, followed by the Gulf of Kachchh and the Gulf of Khambhat. The Gulf of Kachchh supports marine organisms' growth, and it is considered one of the biological most productive marine habitats.
- The habitat preferences and association of intertidal shrimp species were recorded and studied.
- *Alpheus lobidens* is the single species, which was reported from all the microhabitats except mangroves.
- *A. malabaricus* is a single species that was exclusively reported only from the mangrove habitat.
- The variation in the distribution pattern and abundance of organisms in different microhabitats of the intertidal area has provided the basis for so many ecological aspects, especially for the rocky intertidal organisms.
- During the present study, detailed cladistic analyses of prawn and shrimp species were studies based on the 41 morphometric variables.
   For this study, a total of 39 species (28 Dendrobranchiata and 11

Pleocyemata) of prawns and shrimps were used, and its morphometric measurements.

- Dendrobranchiata species had low bootstrap support values for half of the branches and lineages (BS <50%).
- Pleocyemata species had a high bootstrap support value for most branches and lineages (BS>50%).
- In the morphometric analysis species like, *P. merguensis*, *P. japonicus*, and *P. penicillatus* are arranged in the same clade. It also supported the molecular phylogeny data.
- DNA was extracted from 47 different prawn and shrimps collected from other locations and habitats along the Gujarat coast. It is also the first molecular phylogenetic study of prawns and shrimps in Gujarat.
- Most of the generated or developed sequences show the 90-100% similarity with the conspecies database sequences in GeneBank.
- In the present study, a total of 42 sequences (27 sequences of Dendrobranchiata and 15 sequences of Pleocyemata) were developed, and 30 sequences were successfully uploaded on NCBI.
- During this study, 4 sequences were first time submitted on the NCBI database.
- Our molecular phylogeny proposes a close relationship (sister position) between *A. lobidens* and *A. pacificus*

# Conclusion

The morphological examination of all the specimens collected from the Gujarat coast of India resulted in identifying 52 different species of prawn and shrimps belonging to 27 genera, 11 families, and 4 superfamilies. Under Suborder Dendrobranchiata and Pleocyemata, 2 families, 10 genera and 9 families, and 17 genera were identified. Family Penaeidae comprised the maximum number of species diversity (25 species, 9 genera), followed by the family Alpheidae (8 species, 3 genera), Plaemonidae (6 species, 4 genera), Hippolytidae (3 species, 3 genera), Solenoceridae (3 species, 1 genus), Callianassidae (2 species, 2 genera), whereas families Lysmatidae,

Thoridae, Pandaloidea, Upogebiidae and Spongicolidae each comprised with a single species. Among the infraorder Caridea, the superfamily Alpheoidea dominated the number of species (13), representing 25% of the total species. Following the family-wise contribution. The analysis reveals 48% of the species are contributed by the family Penaeidae, followed by the family Alpheidae (15%), Plaemonidae (11%). Families Hippolytidae and Solenoceridae have contributed 6% each of the total species diversity.

Information on species distribution and habitat preference of marine invertebrates is the fundamental requirement to understand the presence of different species in benthic communities, providing baseline information for successful conservation of the habitat and benthic fauna. Studies on the distribution and diversity of local fauna are of great importance because they lead to the best understanding of the local animal community's structure, function, and problems (Fransozo et al., 1992; Hebling et al., 1994). The major highlights and recommendations for future studies are listed below.

The morphometrical analysis was conducted using a truss network system on 28 Dendrobranchiata and 11 Pleocyemata shrimp species collected from the Gujarat coast during the present study. Fifty-one morphometric characters representing the carapace, thoracic appendages, and abdominal segments were used to derive the truss network system's (morphometric matrix). In the taxonomical relationship of 28 shrimp of Dendrobranchiata based on morphometrical characters, the standardized size values of species were closely related to respective genera such as *Penaeus*, *Metapenaeus, Parapenaeopsis, Parapenaeus,* and *Solenocera* clearly explained the species relationship.

UPGMA cluster analysis showed *Metapenaeus* and *Parapenaeopsis* species came closely, whereas *Penaeus monodon* formed a separated clade. Morphometric relationships of penaeidae were congruent with the molecular and morphological classification of previous studies. Based on the results, it has been concluded that discriminant function analysis and cluster analysis proved to be an effective procedure for distinguishing and classifying species and describing the taxonomical relationship of penaeidae species. *P. penicillatus, P. japonicus, P. merguiensis,* and *P. cancaliculatus* came closely. Species belonging to the family solenoceridae are close to each other, and they are not arranged in a single clade except one species *viz. S. choprai. P. indicus, P. merguiensis,* and *P. penicillatus* are morphologically similar, which was also supported by the cluster analysis. All the species belonging to the genus *Metapenaeus* came closely except three viz *M. kutchensis, M. moyebi,* and *M. dobsoni.* In the suborder pleocymeta, the UPGMA cluster analysis based on the morphometric traits does not work effectively. For this, we need a greater number of specimens of all the families that come under this group.

The phylogenetic study of Dendrobranchiata and Pleocymeta species was investigated based on COI gene sequencing analysis collected from the Gujarat coast during the present. DNA was successfully extracted from the 47 samples, and 42 samples were successfully amplified using the two different primers. The positive amplicon was sequenced, and a total 42 species sequences were developed. The BLASTn analysis of all 42 sequences was performed and 70% of species clearly matched with respective species available on the NCBI database. In addition, 6 sequences were obtained from the NCBI for the phylogenetic analysis. The phylogenetic analysis of Dendrobranchiata involved 28 nucleotide sequences (27 developed and 1 obtained). It is well supported. All the species level grouping within the genus *Alpheus* came under a subsequent clade that was strongly supported with high bootstrap value. The position of the family Alpheidae closer to Hippolytidae. Based on the cladistic analysis, the taxonomy of Hippolytidae is not clear.

The standardized usage of mt COI gene sequences as DNA barcoding has emerged as an accurate tool for the rapid identification of various organisms providing high species resolution (e.g., Costa et al., 2007; Burns et al., 2008) and is increasingly used for the prawn identification (Mamatha et al., 2016; Subbaiya et al., 2017; Kundu et al., 2018). The use of the mitochondrial COI gene proves to be a useful technique in resolving longstanding problems in the identification of prawns. Morphological examination of cryptic species can lead to incorrect identification (e.g., genus *Alpheus*; on the other and no misidentification occurs using DNA barcoding. The morphological examination approach overlooks morphologically cryptic species. The standard method described in this chapter can easily be applied and used anywhere else to identify prawn and shrimp. Our study presents the most comprehensive and robust molecular phylogenetic survey of Dendrobranchiata and Pleocymeta species to date from the Gujarat coast. It is also the first molecular phylogenetic study.

In the present study, the phylogenetic relationship of different genera *Penaeus, Metapenaeus, Parapenaeopsis, Penaeopsis,* and the other genera reported based on the molecular tool is congruent with cluster analysis. Thus, it could be concluded that molecular phylogeny using the mtCOI gene is more useful for phylogenic studies of Dendrobranchiata and Pleocymeta species than traditional morphometric study.

#### Recommendation

- The present study showed that the coastal area of the Gujarat state supports a huge diversity of prawn and shrimps, and detail taxonomic studies are still required to carve a clear picture of the intertidal shrimp fauna of Gujarat.
- The detailed studies on intertidal zonation patterns of marine invertebrates are still required for intertidal shrimps' symbiotic study.
- The coastal areas of Gujarat state support different kinds of marine habitats like the rocky shore, muddy with a rocky bottom, and mudflat was classified as a macro-habitat. In the present study, the micro-habitat pattern was established only based on the dominant animal community present. Future studies can be carried out based on the zonation pattern of shores.

- The rocky shore is the most productive shoreline in terms of diversity, and a detailed study of rocky shore habitat in terms of ecology is still required.
- In the case of caridean shrimps' study of specialized habitats *e.g.*, benthic ground, uniform vegetated canopy and color of host species are required. These habitats can promote the subsequent selection of several adaptive traits, such as camouflage involving remarkable shape and color changes.
- In the morphometric study, we have examined the smaller number of species from Gujarat state. Additional data needed to revise some caridean families' taxonomy like Palaemonidae, Hippolytidae s.l. and Pasiphaeidae, and such taxonomic revision needs additional taxonomic sampling (worldwide) to resolve the taxonomic relationship between these families.
- A limited number of species were examined (each genus and family) in the present study. A complete mitochondrial genome of these groups must understand the comprehensive phylogeny of the Dendrobranchiata and Pleycometa.

# References

Abele LG and Felgenhauer BE (1986). Phylogenetic and phenetic relationships among the lower Decapoda. *Journal of Crustacean Biology*, 6(3): 385-398.

Achuthankutty CT and George MJ (1973). *Acetes sibogalis* sp. nov. (Crustacea: Decapoda: Sergestidae) from Cochin backwaters with a note on its impregnation. *Indian journal of marine sciences*, 2(2): 139-144.

Achuthankutty CT and Nair S (1976). A new species of sergestid shrimp *Acetes orientalis* (Crustacea: Decapoda: Sergestidae) from Goa, Central West Coast of India. *Hydrobiologia*, 48(3): 233-239.

Achuthankutty CT and Nair SRS (1993). A new record of two penaeid species from Goa Coast. *Journal of the Indian Fisheries Association*, 23: 109-111.

Achuthankutty M and Murthy BVSR (1991). On a new record of *Metapenaeopsis toloensis* Hall (Crustacea: Decapoda) from Indian waters. *Journal of Marine Biological Association of India*, 33: 423-425.

Ahmad E (1972). Coastal Geomorphology of India. Orient Longman, New Delhi, 222 pp.

Akash S, Purushothaman P, Madhavan M, Ravi C, Hisham TJ, Sudhakar M, Kumar TTA and Kuldeep LK (2020). *Urocaridella arabianensis* n. sp., a new Palaemonid shrimp (Crustacea, Decapoda, Palaemonidae) from Lakshadweep Islands, India with taxonomic comparison on the genus *Urocaridella* Borradaile, 1915. *Zootaxa*, 4816(1): 49-66.

Aktas M, Cemal T and Bozkurt A (2006). Taxonomic description of three shrimp species (*Melicertus kerathurus, Meatapenaeus monoceros, Penaeus semisulcatus*) using multivariate morphometric analyses. Journal of Animal and Veterinary Advances, 5: 172-175.

Alcock A (1901). A descriptive catalogue of the Indian deep-sea Crustacea Decapoda Macrura and Anomala, in the Indian Museum. Being a revised account of the deep-sea species collected by the Royal Indian Marine Survey Ship Investigator. *Trustees of the Indian Museum*: Calcutta, 286 pp.

Alcock A (1905). A revision of the" Genus" *Peneus*, with diagnoses of some new species and varieties. *The Annals and Magazine of Natural History*, (7)16: 508-532.

Alcock A (1906). Catalogue of the Indian Decapod Crustacea in the Collection of the Indian Museum: Macrura Part 3; The Prawns of the *Peneus* Group. The Trustees of the Indian Museum, Calcutta, 57 pp. Alcock A and Anderson ARS (1894). Natural history notes from H.M. Royal Indian marine survey steamer 'Investigator', Commander C.F. Oldham, R.N., commanding. Series II, no. 14. An account of a recent collection of deep-sea Crustacea from the Bay of Bengal and Laccadive Sea. *Journal of the Asiatic Society of Bengal*, 63: 141-185.

Alcock A and Anderson ARS (1899). Natural history notes from H.M. Royal Indian marine survey ship'Investigator', Commander T.H. Heming, R.N., commanding. - Series III, no. 2. An account of the deep-sea Crustacea dredged during the surveying-season of 1897-98. *The Annals and Magazine of Natural History*, (7) 3: 1-27, 278-292.

Almeida AO, Terossi M and Mantelatto FL (2014). Morphology and DNA analyses reveal a new cryptic snapping shrimp of the *Alpheus heterochaelis* Say, 1818 (Decapoda: Alpheidae) species complex from the western Atlantic. *Zoosystema*, 36 (1): 53-71.

Alvarez F, Iliffe TM and Villalobos JL (2006). Macromaxillocarididae, a new family of stenopodidean shrimp from an anchialine cave in the Bahamas, with the description of *Macromaxillocaris bahamaensis*, n. gen., n. sp. *Journal of Crustacean Biology*, 26: 366-378.

Anker A and De Grave S (2016). An updated and annotated checklist of marine and brackish caridean shrimps of Singapore (Crustacea, Decapoda). *Raffles Bulletin of Zoology*, 34: 343-454.

Anker A, Hur C and Knowlton N (2009). Description of cryptic taxa within the *Alpheus bouvieri* A. Milne-Edwards, 1878 and *A. hebes* Kim and Abele, 1988 species complexes (Crustacea: Decapoda: Alpheidae). *Zootaxa*, 2153: 1-23.

Avise JC, Arnold J, Ball RM, Bermingham E, Lamb T, Neigel JE, Reeb CA and Saunders NC (1987). Intraspecific phylogeography: the mitochondrial DNA bridge between population genetics and systematics. *Annual review of ecology and systematics*, 18: 489-522. Avise JC, Giblin-Davidson C, Laerm J, Patton JC and Lansman RA (1979). Mitochondrial DNA clones and matriarchal phylogeny within and among geographic populations of the pocket gopher, *Geomys pinetis*. *Proceedings of the National Academy of Sciences*, 76(12): 6694-6698.

Aznar-Cormano L, Brisset J, Chan TY, Corbari L, Puillandre N, Utge J, Zbinden M, Zuccon D and Samadi S (2015). An improved taxonomic sampling is a necessary but not sufficient condition for resolving interfamilies relationships in Caridean decapods. *Genetica*, 143(2): 195-205.

Baby ST, Ghosh S, Mohan G, Cubelio SS and Sudhakar M (2015). Occurrence of Marbled Shrimp *Saron marmoratus* (Olivier, 1811) (Decapoda: Caridea: Hippolytidae) in Lakshadweep Archipelago, India. *Proceedings of the Zoological Society*, 69(1): 157-160.

Balss H (1957). Decapoda. VIII. Systematik. In: Dr. H.G Bronns' Klassen und Ordnungen des Tierreichs. Funfter Band, 1. Abteilung 7, Buch 12. Winter, Leipzig/Heidelberg, 1505-1672 pp.

Banner AH and Banner DM (1972). The establishment of a neotype for *Alpheus edwardsi* (Audouin). *Bulletin du Muséum national d'Histoire Naturelle, Paris series,* 3 (88): 1141- 1146.

Banner AH and Banner DM (1979). Some small collections of alpheid shrimp from the Indian Ocean, including two new species of the genus *Synalpheus*. *Pacific Science*, 33: 25-35.

Banner DM and Banner AH (1982). The alpheid shrimp of Australia. Part III: The remaining alpheids, principally the genus *Alpheus* and the family Ogyrididae. *Records of the Australian Museum*, 34: 1-357.

Bate CS (1881). On the Penaeidae. *The Annals and Magazine of Natural History, (series 5)* 8: 169-196.

Bate CS (1888). Report on the Crustacea Macrura collected by the Challenger during the years 1873-76. *In Report on the Scientific Results of the Voyage of H.M.S. Zoology*, 24 (part 52): 1-942.

Bauer RT (2004). Remarkable Shrimps: Adaptations and Natural History of the Carideans. University of Oklahoma Press, USA, 282 pp.

Bektas Y and Belduz AO (2009). Morphological variation among atlantic horse mackerel, Trachurus trachurus populations from Turkish coastal waters. *Journal of Animal and Veterinary Advances*, 8(3): 511–517.

Beleem I, Poriya P and Gohil B (2019). First record of the callianassid ghost shrimp *Neocallichirus jousseaumei* (Nobili, 1904) (Decapoda: Axiidea) from India. *Journal of Threatened Taxa*, 11(3): 13402-13405.

Bharathi S, Purushothaman P, Akash S, Jose S, Madhavan M, Dhinakaran A, Saravanane N, Kumar TTA and Lal KK (2019). *Periclimenella agattii* sp. nov., a new Palaemonid shrimp (Crustacea, Decapoda, Palaemonidae) from Lakshadweep Islands, India. *Zootaxa*, 4706(3): 483-493.

Bhat UG and Neelakantan B (1984). Composition and distribution of benthos in Kaliestuary, Karwar. *Journal of the Indian Fisheries Association*, 14(15): 27-35.

Bhuti GS (1976). Studies on the alpheids of the Karwar area (Crustacea, Natantia, Decapoda) (Ph.D. Thesis). *The Karnatak Uniyersity,* Karwar, 1-324 pp.

Bilgin R, Utkan MA, Kalkan E, Karhan SU and Bekbölet M (2015). DNA barcoding of twelve shrimp species (Crustacea: Decapoda) from Turkish seas reveals cryptic diversity. *Mediterranean Marine Science*, 16(1): 36-45.

Bissaro FG, Gomes-Jr JL and Di Beneditto AP (2013). Morphometric variation in the shape of the cephalothorax of shrimp *Xiphopenaeus kroyeri* on the east coast of Brazil. Marine Biological Association of the United

Kingdom. Journal of the Marine Biological Association of the United Kingdom, 93(3): 683-691.

Bock WJ (1974). Philosophical Foundations of Classical Evolutionary Classification. *Systematic Zoology*, 22: 375-392.

Boltana S and Thiel M (2001). Associations between two species of snapping shrimp, *Alpheus inca* and *Alpheopsis chilensis* (Decapoda: Caridea: Alpheidae). *Journal of the Marine Biological Association of the United Kingdom*, 81(4): 633.

Bookstein FL (1991). Morphometric tools for landmark data: Geometry and Biology. Cambridge University press, Cambridge, 1-435 pp.

Borradaile LA (1907). On the classification of the decapod crustaceans. *Annals and Magazine of Natural History*, 7(19): 457-486.

Bouvier EL (1917). Crustace<sup>°</sup>is decapods (*Macroures marcheurs*) provenant des campagnes des yachts Hirondelle et Princesse-Alice (1885-1915). *Campagnes Scientifiques Monaco*, 50: 1-140.

Bracken HD, De Grave S and Felder DL (2009a). Phylogeny of the infraorder Caridea based on mitochondrial and nuclear genes (Crustacea: Decapoda). *Decapod crustacean phylogenetics*, 18: 274-298.

Bracken HD, Toon A, Felder DL, Martin JW, Finley M, Rasmussen J, Palero F and Crandall KA (2009b). The decapod Tree of Life: compiling the data and moving toward a consensus of decapod evolution. *Arthropod Systematics and Phylogeny*, 67 (1): 99-116.

Bracken HD, De Grave S, Toon A, Felder DL and Crandall KA (2010). Phylogenetic position, systematic status, and divergence time of the Procarididea (Crustacea: Decapoda). *Zoologica Scripta*, 39(2): 198-212.

Bracken-Grissom HD, Robles R and Felder DL (2014). Molecular phylogenetics of American snapping shrimps allied to *Alpheus floridanus* 

Kingsley, 1878 (Crustacea: Decapoda: Alpheidae). *Zootaxa*, 3895(4): 492-502.

Bremner J, Rogers SI and Frid CLJ (2003). Assessing functional diversity in marine benthic ecosystems: a comparison of approaches. *Marine Ecology Progress Series*, 254: 11-25.

Brown WM, Prager EM, Wang A and Wilson AC (1982). Mitochondrial DNA sequences of primates: tempo and mode of evolution. *Journal of molecular evolution*, 18: 225-239.

Bruce AJ (1995). *Latreutes anoplonyx* Kemp, 1914 (Crustacea: Decapoda: Hippolytidae), a jelly-fish associate new to the Australian fauna. *The Beagle, Records of the Museums and Art Galleries of the Northern territory*, 12: 61-64.

Bruce AJ and Svoboda A (1983). Observations upon some pontoniine shrimps from Aqaba, Jordan. *Zoologische Verhandelingen* 205: l-44.

Burkenroad MD (1934). Littoral Penaeidea chiefly from the Bingham oceanographic collection, with a revision of *Penaeopsis* and descriptions of two genera and eleven new American species. *Bulltetin of Bingham Oceanographic Collection*, 4: 1-109.

Burkenroad MD (1983). A natural classification of the Dendrobranchiata, with a key to recent genera. *Crustacean Phylogeny, Crustacean*, 1: 279-290.

Burns JM, Janzen D H, Hajbabaei M, Hallwachs W and Hebert PDN (2008). DNA barcodes and cryptic species of skipper butterflies in the genus *Perichares* in area de conservacion Guanacaste. *Proceedings of the National Academy of Sciences*, 105: 6350-6355.

Burukovsky RN (1972). Some problems of the systematics and distribution of shrimps of the genus Penaeus [in Russian]. *Trudy AtlantNIRO, Kaliningrad* 2: 3-21.

Çakmak E and Alp A (2010). Morphological differences among the Mesopotamian spiny eel, *Mastacembelus mastacembelus* (Banks & Solander 1794), populations. *Turkish Journal of Fisheries and Aquatic Sciences*, 10: 87-92.

Calado R (2008). Marine ornamental shrimps: biology, aquaculture and conservation. Oxford: Wiley International, United Kingdom, 1-263 pp.

Calman WT (1909). Crustacea. In: Lankester, R., A Treatise on Zoology, Part 7, 3rd Fascicle 3. Adam and Charles Black, London, 196-223 pp.

Calo-Mata P, Pascoal A, Fernández-No I, Böhme K, Gallardo JM and Barros-Velázquez J (2009). Evaluation of a novel 16S rRNA/tRNAVal mitochondrial marker for the identification and phylogenetic analysis of shrimp species belonging to the superfamily Penaeoidea. *Analytical biochemistry*, 391(2): 127-134.

Carvalho CD, Keunecke KA and Lavrado HP (2019). Morphometric variation in pink shrimp populations at Rio de Janeiro coast (SE Brazil): are they really similar in closer areas?. *Anais da Academia Brasileira de Ciências*, 91(2): e20180252.

Carvalho FL, De Grave S and Mantelatto FL (2017). An integrative approach to the evolution of shrimps of the genus *Palaemon* (Decapoda, Palaemonidae). *Zoologica Scripta*, 46(4): 473-85.

Chace Jr FA (1972). The shrimps of the Smithsonian-Bredin Caribbean Expeditions with a summary of the West Indian shallow-water species (Crustacea: Decapoda: Natantia). *Smithsonian Contributions to Zoology*, 98: 1-179.

Chace Jr FA (1992). On the Classification of the Caridea (Decapoda). *Crustaceana*, 63 (1): 70-80.

Chace Jr FA and Bruce AJ (1993). The caridean shrimps (Crustacea: Decapoda) of the Albatross Philippine expedition 1907-1910, part 6:

superfamily Palaemonoidea. *Smithsonian contributions to zoology*, 543: 1-164.

Chace Jr FA and Manning RB (1972). Two new caridean shrimps, one representing a new family, from marine pools on Ascension Island (Crustacea: Decapoda: Natantia). *Smithsonian Contributions to Zoology*, 131: 1-18.

Chakraborty RD and Thumber BP (2005). Note on a fishery of *Parapenaeus longipes* along the Veraval Coast. *Marine Fisheries Information Service, Technical and Extension Series*, 183: 15.

Chakraborty RD, Purushothaman P, Kuberan G, Sebastian J and Maheswarudu G (2015). Morphological analysis and molecular phylogeny of *Aristeusalcocki* Ramadan, 1938 from south-west coast of India. *Indian Journal of Geo Marine sciences*, 44(11): 1716-1725.

Chan TY (1998). Shrimps and prawns, Lobsters. In: Shrimps and prawns, Lobsters. In: FAO Species identification guide for fisheries purpose. The living marine resources of the Western Central Pacific. Volume 2. Cephalopods, crustaceans, holothurians and sharks. K. E. Carpenter & V. H. Niem (Eds). Food and Agriculture Organization, Rome, 851-1043 pp.

Chan TY and Yu HP (1985). Studies on the shrimps of the genus *Palaemon* (Crustacea, Decapoda, Palaemonidae) from Taiwan. *Journal of the Taiwan Museum*, 8(1): 114-127.

Chan TY, Tong J, Tam YK and Chu KH (2008). Phylogenetic relationships among the genera of the Penaeidae (Crustacea: Decapoda) revealed by mitochondrial 16S rRNA gene sequences. *Zootaxa*,1694: 38-50.

Chanda A (2014). A systematic study on genus *Metapenaeus* wood-mason, 1891 with special reference to extended distribution of four species from Indian water. *International Journal of Fisheries and Aquatic Studies*, 2(1): 5-16.

Chanda A (2015). Indian records of the Genus *Metapenaeopsis* Bouvier, 1905 with special reference to extended distribution of two species. *International Journal of Fisheries and Aquatic Studies*, 2(6): 265-273.

Chanda A (2016). A study on newly described genera *Alcockpenaeopsis*, *Batepenaeopsis*, *Helleropenaeopsis*, *Kishinouyepenaeopsis* and *Parapenaeopsis* from Indian water. *Poultry*, *Fisheries & Wildlife Sciences*, 4(1):1-12.

Chanda A (2018). A taxonomic study on trachypenaeid prawns with special reference to Indian records. *Journal of Threatened Taxa*, 10(2):11332-11338.

Chanda A and Bhattacharya T (2002). *Melicertus similis*, a new species of prawn, Decapoda: Penaeidae, from India. *Journal of the Bombay Natural History Society*, 99: 495-498.

Chanda A and Bhattacharya T (2003). *Fenneropenaeus konkani*, a new species of prawn (Decapoda: Penaeidae) from Indian Coast. *Science and Culture*, 69: 229-230.

Chanda A and Roy T (2004). Prawns of Gujarat Coast. *Zoological Survey of India State Fauna of Gujarat,* 8: 211-218.

Chandra A and Bhattacharya T (2004). A new species of the genus *Parapenaeopsis* Alcock, 1901 (Penaeoidea: Penaeidae) from Orissa, India. *Proceedings of the Zoological Society*, 57: 23-27.

Chavanich S and Wilson KA (2000). Rocky intertidal zonation of gammaridean amphipods in Long Island Sound, Connecticut. *Crustaceana*, 73(7): 835-846.

Chen CL, Goy JW, Bracken-Grissom HD, Felder DL, Tsang LM and Chan TY (2016). Phylogeny of Stenopodidea (Crustacea: Decapoda) shrimps inferred from nuclear and mitochondrial genes reveals non-monophyly of

the families Spongicolidae and Stenopididae and most of their composite genera. *Invertebrate Systematics*, 30(5): 479-490.

Chopra B (1923). Bopyrid isopods parasitic on Indian Decapoda Macrura. Decapoda, Macrura. *Records of Indian museum*, 25: 411-550.

Choudhury A, Bhunia AB and Nandi S (1984). Preliminary survey on macrobenthos of Prentice Island, Sundarbans, West Bengal. *Records of Zoological Survey India*, 81 (3 & 4): 81-92.

Chowdhury LM, Kathirvelpandian A, Divya PR, Basheer VS, Shanis R, Chelath M, Kumar AP and Krishna G (2019). Molecular identification and phylogenetic assessment of species under genus *Parapenaeopsis* Alcock, 1901, from Indian waters. *Mitochondrial DNA Part A*, 30(2): 191-200.

Christoffersen ML (1986). Phylogenetic relationships between Oplophoridae, Atyidae, Pasiphaeidae, Alvinocarididae fam. n., Bresiliidae, Psalidopodidae and Disciadidae (Crustacea Caridea Atyoidea). *Boletim de Zoologia, Universidade de São Paulo,* 10: 273-281.

Christoffersen ML (1987). Phylogenetic relationships of hippolytid genera, with an assignment of new families for the Crangonoidea and Alpheoidea (Crustacea, Decapoda, Caridea). *Cladistics*, 3: 348-362.

Christoffersen ML (1988a). Genealogy and phylogenetic classification of the world Crangonidae (Crustacea, Caridea), with a new species and new records for the South Western Atlantic. *Revista Nordestina de Biologia*, 6(1): 43-59.

Christoffersen ML (1988b). Phylogenetic systematics of the Eucarida (Crustacea, Malacostraca). *Revista Brasiliera de Zoologia*, 5(2): 325-351.

Christoffersen ML (1989). Phylogeny and classification of the Pandaloidea (Crustacea, Caridea). *Cladistics*, 5: 259-274.

Christoffersen ML (1990). A new superfamily classification of the Caridea (Crustacea: Pleocyemata) based on phylogenetic pattern. *Journal of Zoological Systematics and Evolutionary Research*, 28(2): 94-106.

CMFRI (1969). Prawn Fisheries of India. *The Bulletin of the Central Marine Fisheries Research Institute*, 14:1-303.

Convention on Biological Diversity (1992).

Costa FO, Dewaard JR, Boutillier J, Ratnasingham S, Dooh RT, Hajibabaei M and Hebert PDN (2007). Biological identifications through DNA barcodes: the case of the crustacea. *Canadian Journal of Fisheries and Aquatic Sciences*, 64: 272-295.

Costello MJ, Coll M, Danovaro R, Halpin P, Ojaveer H and Miloslavich P (2010). A Census of Marine Biodiversity Knowledge, Resources, and Future Challenges. *PLoS ONE*, 5(8) e12110: 1-15.

Coutière H (1903). Note sur quelques Alpheidæ des Maldives et Laquedives. *Bulletin de la Société Philomathique de Paris*, (9)5: 72-90.

Coutière H (1905). Les Alpheidae. In: Gardiner, J.S., The Fauna and Geography of the Maldive and Laccadive Archipelagoes. Being the account of the work carried on and of the Collections made by an Expedition during the years 1899 and 1900. University Press, Cambridge, 852-921 pp.

Coutière H (1921). The Percy Sladen Trust Expedition to the Indian Ocean in 1905, under the leadership of Mr. J. Stanley Gardiner, M.A. No. X. — Les espéces d'Alpheidæ rapportées de l'Océan Indien par M. J. Stanley Gardiner. *Transactions of the Linnean Society of London* (2) *Zoology*, 17: 413-428.

Dall W (1999). Australian species of Solenoceridae (Penaeoidea: Decapoda). *Memoirs of the Queensland Museum*, 43(2):553-587.

Dall W, Hill BJ, Rothlisberg NW and Staples DJ (1990). The Biology of Penaeidae. *Advances in Marine Biology*, 27:1-484.

Darwin C (1859). On the origin of species by means of natural selection, or the Preservation of favoured races in the struggle for life, 1st Edn. John Murray, London, 45 pp.

Dash G, Sen S, Koya M, Sreenath KR, Mojjada S and Bhint HM (2012). Ginger Prawn Fishery in Gulf of Kutch: A Seasonal Livelihood for the Traditional Fishermen. *Asian Agri-History*, 16(4): 393-401.

De Grave S and Fransen CHJM (2011). *Carideorum catalogus*: The recent species of the dendrobranchiate, stenopodidean, procarididean and caridean shrimps (Crustacea: Decapoda). *Zoologische Mededelingen*, 85: 195-589.

De Grave S, Fransen CH and Page TJ (2015). Let's be pals again: major systematic changes in Palaemonidae (Crustacea: Decapoda). *PeerJ*, 3 (e1167): 1-21.

De Grave S, Li CP, Tsang LM, Chu KH and Chan TY (2014). Unweaving hippolytoid systematics (Crustacea, Decapoda, Hippolytidae): resurrection of several families. *Zoologica Scripta*, 43(5): 496-507.

De Grave S, Pentcheff ND, Ahyong ST, Chan TY, Crandall KA, Dworschak PC, Felder DL, Feldmann RM, Fransen CHJM, Goulding LYD, Lemaitre R, Low MEY, Martin JW, Ng PKL, Schweitzer CE, Tan SH, Shudy D and Wetzer R (2009). A classification of living and fossil genera of decapod crustaceans. *Raffles Bulletin of Zoology, Supplement*, 21: 1-109.

De Man JG (1928). The Thalassinidae and Callianassidae collected by the Siboga-Expedition with some remarks on the Laomediidae. The Decapoda of the Siboga-Expedition. Part VII. Siboga Expedition Monogram, 39a6: 1–187.

De Saint Laurent M (1979a). Vers une nouvelle classification des Crustacés Décapodes Reptantia. *Bulletin del'Office Nationale de Pêche de Tunisie,* 3: 15-31 De Saint Laurent M (1979b). Sur la classification et la phylogénie des Thalassinides: définitions de la superfamilledes Axioidea, de la sous-famille des Thomassiniinae et de deux genres nouveaux (CrustaceaDecapoda). *Comptes Rendus Hebdomadaires de Séances de l'Académie des Sciences, Paris,* 288: 1395-1397.

Deshmukh VD (2006). Fishery and biology of the ginger prawn, *Metapenaeus kutchensis* George, George and Rao, 1963 along the northwest coast of India. *Journal of Marine Biological Association of India*, 48(2): 173-179.

Deshmukh VM (1975). A note on the prawn fishery in the Gulf of Kutch during 1962-63. *Indian Journal of Fisheries*, 22(1 & 2): 265-269.

Diaz RJ and Cutter GR (2001). In situ measurement of organism-sediment interaction: rates of burrow formation/abandonment and sediment oxidation/reduction, in Organism-Sediment Interactions, edited by J. Y. Aller, S. A. Woodin, and R. C. Aller. University of South Carolina Press, Columbia, 19-32 pp.

Dineshbabu AP (2004). An account on the fishery and biology of *Parapenaeus fissuroides indicus* Crosier, 1985, recorded for the first time from Indian waters. *Journal of Marine Biological Association of India*, 46(2):215-219.

Ďuriš Z and Horká I (2017). Towards a revision of the genus *Periclimenes*: resurrection of *Ancylocaris* Schenkel, 1902, and designation of three new genera (Crustacea, Decapoda, Palaemonidae). *ZooKeys*, (646): 25-44.

Ďuriš Z, Horka I, Juračka PJ, Petrusek A and Sandford F (2011). These squatters are not innocent: The evidence of parasitism in sponge-inhabiting shrimps. *PLoS one*, 6(7), e21987: 1-10.

Dworschak PC (2000). Global diversity in the Thalassinidea (Decapoda). *Journal of Crustacean Biology*, 20(5): 238-245.

Dworschak PC (2011). Redescription of *Callianassa jousseaumei* Nobili, 1904, a junior subjective synonym of *Callianassa indica* de Man, 1905 with description of a new species of *Neocallichirus* (Decapoda: Axiidea: Callianassidae). *Zootaxa*, 2746: 1–19.

Elliott NG, Haskard K and Koslow JA (1995). Morphometric analysis of orange roughy (*Hoplostethus atlanticus*) off the continental slope of southern Australia. *Journal of Fish Biology*, 46(2): 202-20.

Ellison JC (2012). Climate Change Vulnerability Assessment and Adaptation Planning for Mangrove Systems. World Wildlife Fund (WWF), Washington, DC, USA, 142 pp.

Fabricius JC (1775). Systema entomologiae, sistens insectorum classes, ordines, genera, species adiectis synonymis, locis, descriptionibus, observationibus. Flensburgi et Lipsiae, 832 pp.

Fabricius JC (1798). Supplementum Entomologiae systematicae. Proft et Storch, Hafniae, Copenhagen, 1-572 pp.

Feldmann RM and Schweitzer CE (2010). The oldest shrimp (Devonian: Fammenian) and remarkable preservation of soft tissue. *Journal of Crustacean Biology*, 30: 629-635.

Felgenhauer BE and Abele LG (1983). Phylogenetic relationships among shrimp-like decapods. *Crustacean Phylogeny, Crustacean,* 1: 291-311.

Fitzgerald DG, Nanson JW, Todd TN and Davis BM (2002). Application of truss analysis for the quantification of changes in fish condition. *Journal of Aquatic Ecosystem Stress and Recovery*, 9(2):115-25.

Folmer O, Black M, Hoeh W, Lutz R and Vrijenhoek R (1994). DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Molecular Marine Biology and Biotechnology*, 3 (5):294–299. Fransen CHJM and De Grave S (2009). Evolution and radiation of shrimplike decapods: an overview. In: Martin J.W., K.A. Crandall and D.L. Felder (eds.), Decapod Crustacean Phylogenetics. CRC Press, Boca Raton, 246-259 pp.

Fransozo A, Negreiros-Fransozo ML and Mantelatto FL (1992). Composicao e distribuicao dos Brachyura (Crustacea, Decapoda) do sublitoralnãoconsolidadonaEnseada da Fortaleza, Ubatuba (SP). *Revista Brasileira de Zoologi*, 52(4): 667-675.

Futuyma DJ (1998). Evolutionary Biology. Sinauer Associates. Inc. Sunderland, 600 pp.

Gagic V, Bartomeus I, Jonsson T, Taylor A, Winqvist C, Fischer C, Slade EM, Steffan-Dewenter I, Emmerson M, Potts SG and Tscharntke T (2015). Functional identity and diversity of animals predict ecosystem functioning better than species-based indices. *Proceedings of the Royal Society B: Biological Sciences*, 282(1801): 1-8.

Gaston KJ and Spicer JI (2004). Biodiversity: an introduction. 2nd edn. Blackwell Publishing, Oxford, UK. Gould, S. J. (1991). Bully for brontosaurus: reflections in natural history. Hutchinson Radius, London, UK

George MJ (1972). Zoogeographic distribution of Indian Penaeidae. *Indian journal of marine sciences*, 1(1): 89-93.

George MJ (1979). Taxonomy of Indian prawns (Penaeidae, Crustacea, Decapoda) In: Kurian, C.V., Sharma, G.S., Mohandas, A. and Antony, A. (Eds.), Contributions to Marine Sciences Dedicated to Dr. C.V. Kurian on the occasion of his Sixtieth Birthday. Prof. Kurian's Shashtyabdhapoorthi Committee, Cochin. 21-59 pp.

George MJ and Muthu MS (1968a). On the occurrence of *Metapenaeopsis barbata* (De Haan) (Decapoda: Penaeidae) in Indian waters with taxonomic

notes on the genus. *Journal of The Marine Biological Association of India*, 10(2): 286-291.

George MJ and Muthu MS (1968b). *Solenocera waltairensis*, a new species of prawn (Decapoda: Penaeidae) from Indian waters. *Journal of The Marine Biological Association of India*, 10(2): 292-297.

George MJ and Rao PV (1968). A new species of *Metapenaeus* (Decapoda, Penaeidae). *Journal of the Marine Biological Association of India*, 8: 146-151

George MJ and George KC (1964). On the occurrence of the caridean prawn *Thalassocaris lucida* (Dana) in the stomach of *Neothunnus macropterus* (Temminck and Schlegel) from the Arabian Sea. *Journal of the Marine Biological Association of India*, 6 (1): 171-172.

George MJ, Nagappan Nayar K and Mahadevan S (1973). Under Water Observations—On A Collection of Shrimps from The Gulf of Mannar off Tuticorin. *Records of the Zoological Survey of India*, 67: 357 365.

George PC, George MJ and Rao PV (1963). *Metapenaeus kutchensis* sp. nov., a penaeid prawn from the Gulf of Kutch. *Journal of the Marine Biological Association of India*, 5(2): 284-288.

Ghosh S, Mohammed G, Polara JP and Bhint HM (2012). Monsoon fishery of juvenile ginger prawns at Little Rann of Kutch, Gujarat in relation to environmental parameters. *Indian Journal of Fisheries*, 59(1): 23-27.

Gibinkumar TR, Sabu S, Pravin P and Boopendranath MR (2012). Bycatch Characterization of Shrimp Trawl Landings off Southwest Coast of India. *Fishery Technology*, 49: 132-140.

Glaessner MF (1969). Decapoda. *In*: Moore, R.C., *Treastise on Invertebrate Palaeontology Part R Arthropoda 4 Vol II. The* Geological Society of America Inc and The University of Kansa Press, Bouder, Colorado, Lawrence, 399-533 pp. Gopalakrishnan A, Divya PR, Basheer VS, Raja Swaminathan T, Kathirvelpandian A, Bineesh KK, Kumar RG and Jena JK (2012). In Director NBFGR (ed), Macro flora and fauna of the Gulf of Mannar - a checklist. National Bureau of Fish Genetic Resources, Lucknow, U.P., India, 1-127 pp.

Gopalakrishnan P and Raju KV (1987). Marine prawn farming along the coast of the Gulf of Kutch: its prospects and problems. National Symposium on Research and Development in Marine Fisheries *CMFRI Bulletin,* 44 (part 2): 317-321.

Gopalakrishnan P, Raju VK, Pillai KM and Thakkar SR (1985). Some observations on the spawning and larval rearing of the ginger prawn *Metapenaeus kutchensis* George, George and Rao. *Seafood Export Journal*, 17(12): 5-8

Gopalakrishnan P, Raju VK, Pillai KM, Joshi KD, Somaiya VJ and Devmurai HL (1987). Observations on fish and prawn seed resources of the Gulf of Kutch and their utilization in aquaculture. *Fishery Technology*, 24: 5-11.

Gravely FH (1927). Fauna of Krusadai Island in the Gulf of Mannar. Orders Decapoda (except Paguridea) and Stomatopoda. *Bulletin of the Madras Government Museum*, 1(1): 135-155.

Gravely FH (1930). Supplement to the littoral fauna of Krusadai Island in the Gulf of Mannar-The Alpheidae of Krusadai Island. *Bulletin of the Madras Government Museum (New series), Natural history section,* 1(2): 77-79.

Guérin-Méneville FE (1857). Animaux articulés à pieds articulés. Crustacés. In: de la Sagra, R., Histoire Physique, Politique et Naturelle de l'Île de Cuba. Arthus Bertrand, Paris, 1-87 pp.

Guo CC, Hwang JS and Fautin DG (1996). Host selection by shrimps symbiotic with sea anemones: a field survey and experimental laboratory analysis. *Journal of Experimental Marine Biology and Ecology*, 202(2): 165-176.

Gupta SR and Deshmukhe G (2000). Coastal and maritime environments of Gujarat: Ecology and Economics. Gujarat Ecological Society, Vadodara, 150 pp.

Haswell WA (1879). On the Australian species of *Penaeus*, in the Macleay Museum, Sydney. *The Proceedings of the Linnean Society of New South Wales*, 4: 38-44.

Hebert PD, Stoeckle MY, Zemlak TS and Francis CM (2004). Identification of birds through DNA barcodes. *PLoS Biology*, 2(10): 1657-1663.

Hebling NJ, Mantelatto FL and Negreiros-Fransozo ML (1994). Levantamentoe distribuicao de braquiúros e Anomuros (Crustacea, Decapoda) dos sediment ssublitorais da região da IlhaAnchieta, Ubatuba (SP). *Boletim do Instituto de Pesca*, 21: 1-9.

Heller C (1865). Crustaceen. In: Reise der osterreichischen Fregatte Novara um die Erde in den Jahren 1857, 1858, 1859 unter den Befehlen des Commodors B. von WüllerstorfUrbair. *Zoologischer Theil. Kaiserlichköniglichen Hofund Staatsdruckerei, Wien*, 2(3): 1–280

Henderson JR (1893). A contribution to Indian carcinology. *Transactions of the Linnean Society of London Zoology*, (2)5: 325-458.

Hennig W (1950). Grundzuge einer Theorie der Phylogenetischen Seystematic. Deutseher ZeAtralvirlag, Berlin.

Holthuis LB (1955). The recent genera of the caridean and stenopodidean shrimps (Class Crustacea, Order Decapoda Super-section Natantia) with keys for their determination. *Zoologische Verhandelingen*, 26: 1-157.

Holthuis LB (1980). Species catalogue. Volume 1-Shrimps and prawns of the world. An annotated catalogue of species of interest to fisheries. FAO Fisheries Synopsis, 1-271 pp. Holthuis LB (1993). The recent genera of the caridean and stenopodidean shrimps (Crustacea, Decapoda), with an appendix on the order Amphionidacea. Nationaal Natuurhistorisch Museum: Leiden, The Netherlands, 1-328 pp.

Hornell J (1916). Report to the Government of Baroda on the marine zoology of Okhamandal in Kattiawar, pt 1. Williams and Norgate, London, 145-148 pp.

Hualkasina W, Sirimontapornb P, Chotigeata W, Querci J and Phongdara A (2003) Molecular phylogenetic analysis of white prawns species and the existence of two clades in *Penaeus merguiensis*. *Journal of experimental marine biology and ecology*, 296: 1–11.

ICMAM (2002). Geographical Information System for Gulf of Kachchh, Integrated Coastal and Marine Area Management Report.

Jayachandran KV (2005). The biodiversity of palaemonid prawns from Indian Seas. Aspects of Aquatic Biodiversity. Special Publication No. 84. CMFRI, Cochi, 21-28 pp.

Jayachandran KV (2010). Indian palaemonid decapod crustaceans: taxonomic status, research challenges and conservation needs. *Indian Journal of Animal Sciences*, 80 (4): 46-52.

Jayachandran KV and Joseph NI (1989). Palaemonid prawn resources on the south-west coast of India. *Journal of Aquaculture in the Tropics,* 4: 65-76.

Johns GC and Avise JC (1998). A comparative summary of genetic distances in the vertebrates from the mitochondrial cytochrome b gene. *Molecular Biology and Evolution*, 15(11): 1481-1490.

Jones WT, Feldmann RM, Schweitzer CE, Schram FR, Behr R and Hand KL (2014). The first Paleozoic stenopodidean from the Huntley Mountain

Formation (Devonian Carboniferous). *Journal of Paleontology*, 88: 1251-1256.

Jose D, Rozario JV, Benjamin D and Harikrishnan M (2014). Morphological and molecular description for *Glyphocrangon investigatoris* Wood-Mason and Alcock, 1891 emphasizing its phylogenetic relationship. *Mitochondrial DNA Part A*, 27(3): 2053-2057.

Jose S, Purushothaman P, Madhavan M, Akash S, Bharathi S, Dhinakaran A, Kumar TTA and Lal KK (2020). Two new records of hippolytoid shrimps, *Lysmata hochi* Baeza amp; Anker, 2008 (Decapoda: Lysmatidae) and *Lysmata amboinensis* (de Man, 1888) from Lakshadweep Islands, India with taxonomic notes. *Zootaxa*, 4755(2): 353-364.

Joseph A and Soni VC (1990). A study on prawn fishery of the mouth of Gulf of Kutch with special reference to certain biological aspects of some prawn species. *Indian Journal of Fisheries,* 37 (3): 175-182.

Joseph A and Soni VC (1986). Length - weight relationship and relative condition factor of prawn, *Metapenaeus kutchensis* George, George and Rao from Okha. *Indian Journal of Fisheries*, 33(1): 127-129.

Kagwade PV (1978). New record of the penaeid prawn *Metapenaeus kutchensis* from Bombay with a redescription of the species. *Journal of the Marine Biological Association of India*, 20(1&2): 174-176.

Karuppasamy PK, Logeshwaran V, Priyadarshini RS and Ramamoorthy N (2020). Molecular and Systematic Identification of Food Marine Shrimps Using mtCOI Marker from Southeast Coast of India. *Thalassas: An International Journal of Marine Sciences*, 36: 487-495

Karuppasamy PK, Menon NG, Nair KKC and Achuthankutty CT (2006). Distribution and abundance of pelagic shrimps from the deep scattering layer of the eastern Arabian Sea. *Journal of Shellfish Research*, 25(3): 1013-1019.

Kathirvel M, Thirumilu P and Gokul A (2007). Indian penaeid shrimps their biodiversity and economical values. In: Editor-Director, National symposium on Conservation and Valuation of Marine Biodiversity. *Zoological Survey of India*: 161-176

Kazmi QB and Kazmi MA (2010). Biodiversity and Biogeography of caridean shrimps of Pakistan. Higher Education Commission, Karachi, 1-400 pp.

Kemp S (1914). Notes on Crustacea Decapoda in the Indian Museum. V. Hippolytidae. *Records of the Indian Museum*, 10: 81-129.

Kemp S (1915). Crustacea Decapoda. Fauna of the Chilka Lake. *Memoirs of the Indian Museum* 3(5): 199-325.

Kemp S (1915). Fauna of the Chilka Crustacea Decapoda. *Memoirs of the Indian Museum*, 5(3): 199-325.

Kemp S (1916). Further notes on Hippolytidae. Notes on Crustacea Decapoda in the Indian Museum. VII. *Record of the Indian Museum*, 12: 385-405.

Kemp S (1922). Notes on Crustacea Decapoda in the Indian Museum. XV. Pontoniinae. *Record of the Indian Museum*, 24: 113-288

Kemp S (1925). Notes on Crustacea Decapoda in the Indian Museum, XVII. On various Caridea. *Record of the Indian Museum*, 27: 249-343.

Keyse J, Crandall ED, Toonen RJ, Meyer CP, Treml EA and Riginos C (2014). The scope of published population genetic data for Indo-Pacific marine fauna and future research opportunities in the region. *Bulletin of Marine Science*, 90(1): 47-78.

Khedkar GD, Reddy AC, Ron TB and Haymer D (2013). High levels of genetic diversity in *Penaeus monodon* populations from the east coast of India. *Springer Plus*, 2(1): 671.

Kim JN, Choi JH, Oh TY, Choi KH and Lee DW (2011). A New Record of Pandalid Shrimp *Procletes levicarina* (Crustacea: Decapoda: Caridea) from Korean Waters. *Fisheries and aquatic sciences*, 14(4): 399-401.

Kimura M (1980). A simple method for estimating evolutionary rate of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution*, 16:111-120.

Kinoshita K (2002). Burrow structure of the mud shrimp *Upogebia major* (Decapoda: Thalassinidea: Upogebiidae). *Journal of Crustacean Biology*, 22(2): 474-480.

Komai T and Shanis CPR (2011). A new species of the genus *Parastylodactylus* Figueira, 1971 (Crustacea: Decapoda: Caridea: Stylodactylidae) from off Kollam, southwest coast of India. *Zootaxa*, 3140: 60-68.

Komai T, Ravinesh R, Riyas A and Kumar AB (2020). New records of two sponge-associated species of *Upogebia* Leach, 1814 from southern India (Decapoda: Gebiidea: Upogebiidae): *U. hexaceras* (Ortmann, 1894) and *U. nithyanandan* (Sakai, Türkay amp; Al Aidaroos, 2015) resurrected from the synonymy of *U. balmaorum* Ngoc-Ho, 1990. *Zootaxa*, 4747(3): 477-494.

Krausman PR (1999). Some basic principles of habitat use. *Grazing behavior of livestock and wildlife*, 70: 85-90.

Kuberan G, Chakraborty RD and Purushothaman P (2019). A new record of deep-sea shrimp *Glyphocrangon investigatoris* Wood-Mason & Alcock, 1891 (Decapoda: Glyphocrangonidae) from the southeastern Arabian Sea. *Zootaxa*, 4612(4): 566-570.

Kuberan G, Chakraborty RD, Purushothaman P and Maheswarudu G (2018a). Two new records of deep sea shrimp under the genus *Parapontocaris* (Crustacea: Decapoda: Crangonidae) from the southwestern coast of India. *Zootaxa*, 4450(5): 581-584.

Kuberan G, Chakraborty RD, Purushothaman P and Maheswarudu G (2018b). Record of the deep sea shrimp *Pasiphaea alcocki* (Wood-Mason & Alcock, 1891) (Crustacea: Decapoda: Pasiphaeidae) from the southwestern coast of India. *Zootaxa*, 4532(4): 597-600.

Kubo I (1942). Studies on the Japanese Palaemonoid Shrimps, III. Leander. *Journal of the Imperial Fisheries Institute*, 35: 17-85.

Kubo I (1949). Studies on penaeids of Japan and its adjacent waters. *Journal of the Tokyo University of Fisheries*, 36: 178-192.

Kumar CP, John BA, Khan SA, Lyla PS and Jalal KCA (2012). Limit of DNA Barcode in Delineating *Penaeus monodon* and in its developing stages. *Sains Malaysiana*, 41(12): 1527-1533.

Kumar JSY, Raghunathan C and Venkataraman K (2015). A report on some symbiotic shrimps (Crustacea: Decapoda) from the Andaman and Nicobar Islands, India. *Scholars Academic Journal of Biosciences*, 3 (1B): 113-119.

Kumar S, Stecher G, Li M, Knyaz C and Tamura K (2018). MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms. *Molecular Biology and Evolution*, 35:1547-1549.

Kumar SV, Pathak KC, Pednekar P, Raju NSN and Gowthaman R (2006). Coastal processes along the Indian coastline. *Current Science*, 91 (4): 530-536

Kundu S, Rath S, Tyagi K, Chakraborty R and Kumar V (2018). Identification of penaeid shrimp from Chilika Lake through DNA barcoding. *Mitochondrial DNA Part B*, 3(1): 161-165.

Kunju MM (1960). On a new record of five species of Penaeidae (Decapoda: Macrura: Penaeidae) on the West Coast of India. *Journal of the Marine Biological Association of India*, 2(1): 82-84. Kuriyan GK (1951). A note on the eggs and the first stage larva of *Hippolysmata vittata* Stimpson. *Journal of the Bombay Natural History Society*, 50: 416-417

Kurup BM, Rajasree R and Venu S (2008). Distribution of deep sea prawns off Kerala. *Journal of the Marine Biological Association of India*, 50(2): 122-6.

Lalitha DS (1980). Notes on three caridean prawns from Kakinada. *Journal* of the Marine Biological Association of India, 22 (1&2), 169-173.

Lalitha DS (1987). Growth and population dynamics of three penaeid prawns in the trawling grounds off Kakinada. *Indian Journal of Fisheries*, 34(2): 245-264.

Lavery S, Chan TY, Tam YK and Chu KH (2004). Phylogenetic relationships and evolutionary history of the shrimp genus *Penaeus* derived from mitochondrial DNA. *Molecular Phylogenetics and Evolution*, 31(1): 39-49.

Lebour MV (1938). Decapod Crustacea associated with the ascidian *Herdmania*. *Proceedings of the Zoological Society of London*, 108B: 649-653.

Li X and Komai T (2003). Pandaloid shrimps from the northern South China Sea, with description of a new species of *Plesionika* (Crustacea: Decapoda: Caridea). *The Raffles Bulletin of Zoology*, 51: 257-275.

Li X, Bruce AJ and Manning RB (2004). Some palaemonid shrimps (Crustacea: Decapoda) from northern South China Sea, with descriptions of two new species. *The Raffles Bulletin of Zoology*, 52: 513-553.

Liao Y, De Grave S, Ho TW, Ip BH, Tsang LM, Chan TY and Chu KH (2017). Molecular phylogeny of Pasiphaeidae (Crustacea, Decapoda, Caridea) reveals systematic incongruence of the current classification. *Molecular phylogenetics and evolution*, 115: 171-180. Lin FJ, Liu Y, Sha Z, Tsang LM, Chu KH, Chan TY, Liu R and Cui Z (2012). Evolution and phylogeny of the mud shrimps (Crustacea: Decapoda) revealed from complete mitochondrial genomes. *BMC genomics*, 13(1):1-2.

Linnaeus C (1758). Systema naturae per regna triae naturae, secundum classis, ordines, genera, species cum characteribus, differentiis, synonyms locis; Edito decima, reformata Vol. 1, 824 Stockholm: Laurentii Salvii.

Ma KY, Chan TY and Chu KH (2009). Phylogeny of penaeoid shrimps (Decapoda: Penaeoidea) inferred from nuclear protein-coding genes. *Molecular Phylogenetics and Evolution*, 53(1): 45-55.

MacArthur RH (1972). Geographical ecology: patterns in distribution of species. New York: Harper and Row. Mantelatto, F. L., Fabíola C. R. F., Biagi, Rand Melo. G. A. S. 2004. Majoid Crabs Community (Crustacea: Decapoda) from Infralittoral Rocky/sandy Bottom of Anchieta Island, Ubatuba. *Brazilian Archives of Biology and Technology*, 47 (2): 273-279.

Mahapatro D, Panigrahy RC, Panda S and Mishra RK (2015). Checklist of intertidal benthic macrofauna of a brackish water coastal lagoon on east coast of India: The Chilika Lake. *International Journal of Marine Science*, 5(33): 1-13.

Mamatha DM, Jyothi S, Sharmila S and Khedkar GD (2016). Molecular phylogenyof South Indian prawn species by DNA barcoding using COI gene as a marker. *Computational Science, Mathematics and Biology,* 1: 56-58.

Mandal S and Harkantra SN (2013). Changes in the soft bottom macrobenthic diversity and community structure from the ports of Mumbai, India. *Environmental Monitoring and Assessment*, 185(1): 653-672.

Martin JW and Davis GE (2001.) An updated classification of the Recent Crustacea. *Los Angeles: Natural History Museum of Los Angeles County*, 9: 1-124.

Mathews LM and Anker A (2009). Molecular phylogeny reveals extensive ancient and ongoing radiations in a snapping shrimp species complex (Crustacea, Alpheidae, *Alpheus armillatus*). *Molecular Phylogenetics and Evolution*, 50(2): 268-281.

Menon PG and Williamson DI (1971). Decapod Crustacea from the International Indian Ocean Expedition the species of *Thalassocaris* (Caridea) and their larvae. *Journal of Zoology*, 165(1): 27-51.

Miers EJ (1878). Notes on the Penæidæ in the collection of the British Museum, with descriptions of some new species. *Proceedings of the Zoological Society of London,* 46(1): 298-310.

Millennium Ecosystem Assessment (2005). Ecosystem sand human wellbeing: synthesis. Washington, DC: Island Press.

Milne Edwards H (1830). Description des genres Glaucothoe, Sicyonie, Sergeste et Acete, de l'ordre des Crustacés Décapodes. *Annales des Sciences Naturelles,* (1) 19: 333-352.

Milne Edwards H (1834-1840). Histoire naturelle des Crustacés, comprenant l'anatomie, la physiologie et la classification de ces animaux. *Librairie encyclopédique de Roret, Paris,* 3: 1-468, 1-532, 1-638, 1-32.

Miquel JCE (1982). Le genre Metapenaeus (Crustacea, Penaeidae): taxonomie, biologie et pêches mondiales. *Zoologische Verhandelingen*, 195: 6-132.

Mishra PS, Chaudhari A, Krishna G, Kumar D and Lakra WS (2009). Genetic diversity in *Metapenaeus dobsoni* using RAPD analysis. *Biochemical genetics*, 47(5-6): 421-426.

Mohamed KH and Rao PV (1971). Estuarine phase in the life history of the commercial prawns of the west coast of India. *Journal of the Marine Biological Association of India*, 13(2): 149-161.

Mojekwu TO and Anumudu CI (2015). Advanced techniques for morphometric analysis in fish. *Journal of Aquaculture Research & Development*, 6(8):1-6.

Monod T and Forest J (2012). A history of crustacean classification. *In Treatise on Zoology-Anatomy, Taxonomy, Biology. The Crustacea*, 3: 403-444.

Muthu MS (1972a). *Parapenaeopsis indica* sp. nov. (Decapoda: Penaeidae) from the Indian waters. *Indian Journal of Fisheries*, 16: 174-180.

Muthu MS (1972b). Taxonomic notes on the penaeid prawn *Metapenaeopsis gallensis* (Pearson, 1905). *Journal of The Marine Biological Association of India*, 14(2): 564-567.

Myers N, Mittermeier RA, Mittermeier CG, da Fonseca GA and Kent J (2000). Biodiversity hotspots for conservation priorities. *Nature*, 403: 853-858.

Naderloo R and Türkay M (2012). Decapod crustaceans of the littoral and shallow sublittoral Iranian coast of the Persian Gulf: Faunistics, Biodiversity and Zoogeography. *Zootaxa*, 3374: 1-67.

Naik RM, Murthy MS, Mansuri AP, Rao YN, Pravez R, Mundkur T, Krishnans, Faidu PJ and Krishna TSVR (1991). Coastal marine ecosystems and anthropogenic pressure in the Gulf of Kachchh. WWF, India and Department of Biosciences, Saurashtra University, Rajkot, 287pp.

Natarajan SA (1942). A note on the prawn fauna of Travancore. *Current Science*, 11(12): 468-469.

Ngoc-Ho N (1995). Une espèce nouvelle de Neocallichirus aux îles Tuamotu, Polynésie française (Crustacea, Decapoda, Thalassinidea). Bulletin du Muséum national d'histoire naturelle. Section A, Zoologie, biologie et écologie animals, 17(1-2): 211-218. NIO (2002). Status of Flora and Fauna of Gulf of Kachchh, India. National Institute of Oceanography, Goa, India.

Nobili G (1903). Crostacei di Pondichéry, Mahé, Bomba, etc. *Bollettino dei Musei di Zoologia ed Anatomia Comparata della Regia Università di Torino*, 18(452): 1-24.

Odum HT (1971). Environment, power, and society. New York: Wiley. Odum HT. 1988. Self-organization, tranformity, and information. *Science*, 242: 1132-9.

Ortmann A (1890). Die Decapoden-Krebse des Strassburger Museums, mit besonderer Berücksichtigung der von Herrn Dr. Döderlein bei Japan und bei den Liu-Kiu-Inseln gesammelten und Z. im Strassburger Museum aufbewahrten Formen. I. Theil. Die Unterordnung Natantia (Boas) (Abtheilungen: Penaeidae und Eucyphidea = Caridae der Autoren). Zoologische Jahrbücher. Abtheilung für Systematik, Geographie und Biologie der Thiere, 5: 437-542.

Pachelle PP, Mendes CB and Anker A (2011). The Indo-West Pacific alpheid shrimp *Athanas dimorphus* Ortmann, 1894: first record for Brazil and the western Atlantic. *Nauplius*, 19(1): 89-96.

Palumbi SR and Benzie J (1991). Large mitochondrial DNA differences between morphologically similar penaeid shrimp. *Molecular Marine Biology and Biotechnology*, 1: 27-34.

Pathan DI and Jalihal DR (1997). Proposed taxonomic revision of some important penaeid prawn genera (Crustacea: Decapoda) of Konkan coast (West coast of India). *Journal of the Bombay Natural History Society*, 94(3): 496-514.

Patwardhan A, Ray S and Roy A (2014). Molecular markers in phylogenetic studies-a review. *Journal of Phylogenetics & Evolutionary Biology*, 2(2): 1-9.

Pawar PR (2011). Species diversity of birds in mangroves of Uran (Raigad), Navi Mumbai, Maharashtra, West coast of India. *Journal of Experimental Sciences*, 2: 73-77.

Pérez Farfante I (1969). Western Atlantic shrimps of the genus *Penaeus*. *Fishery Bulletin*, 67 (3): 461-590.

Pérez Farfante I (1987). Revision of the gamba prawn genus *Pseudaristeus*, with description of two new species (Crustacea: Decapoda: Penaeoidea). *Fishery Bulletin*, 85: 311-338.

Pérez Farfante I and Kensley B (1997). Penaeoid and sergestoid shrimps and prawns of the world. Keys and diagnoses for the families and genera. *Mémoires du Muséum National d'Histoire naturelle*, 175: 1-233.

Pillai NK (1954). A note on *Callianassa maxima* M. Edwards, (Decapoda).*Bulletin of the Central Research Institute of the University of Travancor*, (C)3 (1): 23-26.

Pillai NK (1955). Pelagic Crustacea of Travancore. I. Decapod Lariae. *Bulletin of the Central Research Institute series,* c 4 (1): 47-101.

Pillai NK (1962). On the occurrence of *Microprosthema validum* Stimpson in Indian waters *Journal of the Marine Biological Association of India*, 3(1 & 2): 267-269.

Pillai NS and Gopalakrishnan KA (1984). Brief account of 'Gunja Net' fishing of the Kutch backwaters (Gujarat). *Fishery Technology*, 21: 147-149.

Pillai SL and Thirumilu P (2013). Rediscovery of the deep sea shrimp *Glyphocrangon investigatoris* Wood-Mason and Alcock, 1891 from Indian waters. *Journal of the Marine Biological Association of India*, 55(1): 91-93.

Pillai SL, Kizhakudan SJ, Radhakrishnan EV and Thirumilu P (2014). Crustacean bycatch from trawl fishery along north Tamil Nadu coast. *Indian Journal of Fisheries*, 61(2): 7-13. Pillai SV (1966). Some Observations on the Early Larval Stages of *Hippolysmata vittata* (Stimpson). *Journal of the Marine Biological Association of India*, 8(1): 152-158.

Poore GCB (1994). A phylogeny of the families of Thalassinidea (Crustacea: Decapoda) withkeys to the families and genera. *Memoirs of the Museum of Victoria*, 54: 79-120

Porter ML, Perez-Losada M and Crandall KA (2005). Model-based multilocus estimation of decapod phylogeny and divergence times. *Molecular phylogenetics and evolution*, 37(2): 355-369.

Poupin J, Zubia M, Gravier-Bonnet N, Chabanet P and Malay M (2013). Illustrated checklist of the Decapoda at Europa Island. *Western Indian Ocean Journal of Marine Science*, 11(1): 1-25.

Pradhan A (2011). Living on the Margins: Prawn Harvesters from Little Rann of Kutch (An Exploratory Study of Health Status). Mumbai: Centre for Health and Allied Themes, 1-114 pp.

Prakash S and Marimuthu N (2020). Notes on some crinoid associated decapod crustaceans (Crustacea: Decapoda) of Lakshadweep Archipelago, Central Indian Ocean. *Zootaxa*, 4766(1): 86-100.

Prakash S, Babu I, Gopi M, Kumar TA and Balasubramanian T (2011). Discovery of the shrimp *Pycnocaris chagoae* Bruce, 1972 (Decapoda: Caridea: Gnathophyllidae) in the Lakshadweep Archipelago, India. *Zootaxa*, 2998(1): 66-68.

Prakash S, Kumar TA and Subramoniam T (2016). New records of marine ornamental shrimps (Decapoda: Stenopodidea and Caridea) from the Gulf of Mannar, Tamil Nadu, India. *Check List*, 12 (6): 1-6.

Prakash S, Kumar TTA and Subramoniam T (2015). Notes on some Indo-Pacific Caridean shrimps (Crustacea: Decapoda: Caridea: Palaemonidae and Gnathophyllidae) particularly from India. *Zootaxa*, 3914 (4): 456-466. Pravez R, Krishnan K and Mansuri AP (1992). Sex ratios of *Metapenaeus kutchensis* George, George and RAO, 1963 and *Parapenaeopsis sculptilis* (Heller, 1862) in the Gulf of Kachchh, western India. *Pakistan Journal of Marine Sciences*, 1(1): 1-5.

Pravin P and Manohardoss RC (1996). Constituents of low value trawl bycatch caught off Veraval. *Fishries Technology*, 33 (2): 121-123.

Purushothaman P, Chakraborty RD, Kuberan G and Maheswarudu G (2019). Integrative taxonomy of commercially important deep water penaeoid shrimps from India. *Journal of genetics*, 98(12): 1-13.

Purushothaman P, Chakraborty, RD, Maheswarudu G and Kuberan G (2017). New records of *Solenocera barunajaya* Crosnier, 1994 and *Solenocera rathbuni* Ramadan, 1938 (Crustacea: Decapoda: Penaeoidea) from the southwest coast of India. *Zootaxa*, 4341(2): 296-300.

Radhakrishnan EV, Deshmukh VD, Maheswarudu G, Josileen J, Dineshbabu AP, Philipose KK, Sarada PT, Pillai SL, Saleela KN, Chakraborty R, Dash G, Sajeev CK, Thirumilu P, Sridhara B, Muniyappa Y, Sawant AD, Vaidya NG, Johny RD, Verma JB, Baby PK, Unnikrishnan C, Ramachandran NP, Vairamani A, Palanichamy A, Radhakrishnan M and Raju B (2012). Prawn fauna (Crustacea: Decapoda) of India - An annotated checklist of the Penaeoid, Sergestoid, Stenopodid and Caridean prawns. *Journal of Marine Biological Association of India* 54(1): 50-72.

Radhakrishnan EV, Josileen J, Lakshmi SP (2011). Handbook of prawns. Central Marine Fisheries Research Institute, Kochi, 1-125 pp.

Rajakumaran P and Vaseeharan B (2014). Survey on Penaeidae shrimp diversity and exploitation in south east coast of India. *Fisheries and Aquaculture Journal Fisheries and Aquaculture Journal*, 5(3): 1-8.

Rajakumaran P, Vaseeharan B, Jayakumar R and Chidambara R (2014). Conformation of phylogenetic relationship of Penaeidae shrimp based on morphometric and molecular investigations. *Cytology and genetics*, 48(6): 357-363.

Rajkumar G, Bhavan PS, Udayasuriyan R and Vadivalagan C (2015). Molecular identification of shrimp species, *Penaeus semisulcatus*, *Metapenaeus dobsoni*, *Metapenaeus brevicornis*, *Fenneropenaeus indicus*, *Parapenaeopsis stylifera* and *Solenocera crassicornis* inhabiting in the coromandel coast (Tamil Nadu, India) using MT-COI gene. *International Journal of Fisheries and Aquatic Studies*, 2(4): 96-106.

Ramakrishna SJ and Talukdar S (2003). Marine invertebrates of Digha coast and some recommendations on their conservation. *Records of the Zoological Survey of India*, 101(3-4), 1-23.

Ramamurthy S (1963a). A note on the prawn fishery at Adesar Camp. *Journal of the Marine Biological Association of India*, 5(2): 318-319.

Ramamurthy S (1963b). A note on prawn fishery of Kutch. *Journal of Marine Biological Association of India,* 5: 146-148.

Ramamurthy S (1964). On a new record of *Metapenaeus stebbingi* Nobili, in Indian waters. *Journal of the Marine Biological Association of India*, 6(1): 170-171.

Ramamurthy S (1967). Studies on the prawn fishery of Kutch. *Proceedings of Symposia on Crustacea, Marine Biological Association of India*, 4: 1424-1436.

Rao GS (1983). Observations on the seasonal prawn fishery of the little Rann of Kutch during 1980. *Indian journal of Fisheries*, 30 (1): 24-134.

Rao GS (1984). On a collection of two species of pelagic penaeids (Crustacea: Decapoda) from the oceanic waters of the south-west Arabian Sea. *Journal of Marine Biological Association of India*, 26(1&2): 165-166.

Rao PV and Kartha KNR (1966). On the occurrence of *Callianassa (Callichirus) audax* De Man (Crustacea Decapoda Callianassidae) on the southwest coast of India with a description of male. *Marine Biological Association of India, Symposium, Cochin, Abstracts,* 1:279-284.

Rath S, Dev Roy MK and Ghosh B (2016). Penaeid and Palaemonid Prawns of Godavari Estuary, Andhra Pradesh with Some New Records. *Biological Forum- An International Journal* 8(1): 179-189.

Ravindranath K (1982). The Krishna estuarine complex with reference to its shrimp and prawn fishery. *Indian Journal of* Fisheries, 29: 168-176.

Reddy KN (1995). Crustacea: Decapoda. Fauna of Chilka Lake. Wetland Ecosystem Series, 1 Zoological Survey of India, Calcutta, 367-389 pp.

Reist JD (1985). An empirical evaluation of several univariate methods that adjust for size variation in morpometric data. *Canadian Journal of Zoology*, 63: 1429-1439.

Rönnbäck P, Kautsky N, Pihl L, Troell M, Söderqvist T and Wennhage H (2007). Ecosystem goods and services from Swedish coastal habitats: identification, valuation, and implications of ecosystem shifts. *AMBIO: a Journal of the Human Environment*, 36(7): 534-544.

Saad YM and El-Sebaie HE (2017). The efficiency of cytochrome oxidase subunit 1 gene (cox1) in reconstruction of phylogenetic relations among some crustacean species. *World Academy of Science, Engineering and Technology International Journal of Animal and Veterinary Sciences,* 11(7): 515-20.

Sakai K and Shinomiya S (2011). Preliminary report on eight new genera formerly attributed to *Parapenaeopsis* Alcock, 1901, sensulato (Decapoda, Penaeidae). *Crustaceana*, 84: 491-504.

Salgado-Barragán J, Ayón-Parente M and Zamora-Tavares P (2017). New records and description of two new species of carideans shrimps from

Bahía Santa María-La Reforma lagoon, Gulf of California, Mexico (Crustacea, Caridea, Alpheidae and Processidae). *ZooKeys*, (671): 131-153.

Samuel VKD, Sreeraj CR, Krishnan P, Parthiban C, Sekar V, Chamundeeswari K, Immanuel T, Shesdev P, Purvaja R and Ramesh R (2016). An updated checklist of shrimps on the Indian coast. *Journal of Threatened Taxa*, 8(7): 8977-8988.

Sankarankutty C (1962). On the occurrence of *Athanas dorsalis* (Stimpson) (Decapoda-Alpheidae) in the Gulf of Mannar. *Journal of the Marine Biological Association of India,* 4(2): 167-171.

Sankolli KN (1970). The Thalassinoidea (Crustacea, Anomura) of Maharashtra. *Journal of the Bombay Natural History Society*, 67(2): 235-249.

Sankolli KN (1972). The Thalassinoidea (Crustacea, Anomura) of Maharashtra. *Journal of the Bombay Natural History Society*, 68 (3): 671-682.

Sankolli KN and Shenoy S (1979). On a new genus and a new species of a subterranean prawn *Troglindicus phreaticus* (Caridea, Palaemonidae). *Bulletin of the Fisheries Faculty, Konkan Agricultural University, India,* 1:83-91.

Saravanakumar A, Serebiah JS, Thivakaran GA and Rajkumar M (2007). Benthic macrofaunal assemblage in the arid zone mangroves of Gulf of Kachchh-Gujarat. *Journal of Ocean University of China*, 6(3): 303-309.

Sarkar RJ and Talukdar S (2003). Marine invertebrates of Digha coast and some recommendations on their conservation. *Records of the Zoological Survey of India*, 101: 1-298.

Sarvaiya RT (1981). Prawn fishery of Kutch with special reference to Sukhper and Lakhpat. *Indian journal of Fisheries*, 25 (1&2): 35-40.

Sastry DRK (1977). On some crustacean associates of sea-urchins of the Andaman and Nicobar Islands: Newsletter. *Zoological Survey of India*, 3: 119-120.

Sastry DRK (1981). On some crustacean associates of Echinodermata from the Bay of Bengal. *Records of the Zoological Survey of India*, 79: 19-30.

Sathiya U and Valarmathi V (2018). Diversity of marine shrimp species along the coast of Nagapattinam Tamil Nadu, India. *International Journal of Zoology and Applied*, 3(3): 410-416.

Schram FR (1986). Crustacea. Oxford University Press, New York, New York, 1-606 pp.

Seitz RD, Wennhage H, Bergström U, Lipcius RN and Ysebaert T (2014). Ecological value of coastal habitats for commercially and ecologically important species. *ICES Journal of Marine Science*, 71(3): 648-665.

Shanis CPR, Akhilesh KV, Manjebrayakath H, Ganga U and Pillai NGK (2012). Shrimps of the family Pandalidae (Caridea) from Indian waters, with new distributional record of *Plesionika adensameri* (Balss, 1914). *Journal of the Marine Biological Association of India*, 54(1): 45-49.

Shen X, Li X, Sha Z, Yan B and Xu Q (2012). Complete mitochondrial genome of the Japanese snapping shrimp *Alpheus japonicus* (Crustacea: Decapoda: Caridea): Gene rearrangement and phylogeny within Caridea. *Science China Life Sciences*, 55(7): 591-8.

Silas EG and Muthu MS (1976). On a new species of penaeid prawn of the genus *Metapenaeus* Wood Mason and Alcock, from the Andamans. *Journal of the Marine Biological Association of India,* 16: 645-648.

Simon C, Frati F, Beckenbach A, Crespi B, Liu H and Flook P (1994). Evolution, weighting, and phylogenetic utility of mitochondrial gene sequences and a compilation of conserved polymerase chain reaction primers. *Annals of the Entomological Society of America*, 87:651-701 Solomon SG, Okomoda VT and Ogbenyikwu AI (2015). Intraspecific morphological variation between cultured and wild *Clarias gariepinus* (Burchell) (Clariidae, Siluriformes). *Archives of Polish Fisheries*, 23(1): 53-61.

Stebbing TR (1902). South African Crustacea. Part II. Marine Investigations in South Africa. Cape of Good Hope, Department of Agriculture, Capetown, 1-91 pp.

Strauss RE and Bookstein FL (1982). The truss: body form reconstructions in morphometrics. *Systematic Biology*, 31(2): 113-35.

Subba Rao NY and Sastry DRK (2005). Fauna of Marine National Park, Gulf of Kachchh (Gujarat): An Overview. *Conservation Area Series*, 23: 1-79.

Subbaiya R, Narasimman S, Lawanya R, Mahavinod Angrasan JK, Masilamani Selvam M (2017). Studies on proximal composition and DNA barcoding of marine shrimps from Thondi, Tamilnadu, India. *International Research Journal of Pharmacy*, 8: 59-62.

Subrahmanyam M (1967). An abnormal form of *Alpheus malabaricus* Fabricus from the Godavari estuarine system. *Journal of the Marine Biological Association of India*, 9 (1): 201-202.

Sun MM, Huang JH, Jiang SG, Yang QB, Zhou FL, Zhu CY, Yang LS and Su TF (2014). Morphometric analysis of four different populations of *Penaeus monodon* (Crustacea, Decapoda, Penaeidae). *Aquaculture Research*, 45: 113-23.

Suseelan C (1990). Occurrence of *Heterocarpus sibogae* De Man and *Plesionika williamsi* Forest (Crustacea, Decapoda, Pandalidae) in the Arabian Sea. *Indian Journal of Fisheries*, 37 (4): 321-326.

Suseelan C (1996). Crustacean biodiversity, conservation and management. In: Marine Biodiversity Conservation and Management. (Eds. NG Menon & CSG Pillai), CMFRI, Kochi, 41-64 pp.

Tamaki A (1988). Effects of the bioturbating activity of the ghost shrimp *Callianassa japonica* Ortmann on migration of a mobile polychaete. *Journal of Experimental Marine Biology and Ecology*, 120: 81-95.

Tavares C, Serejo C and Martin JW (2009). A preliminary phylogenetic analysis of the Dendrobranchiata based on morphological characters. In: Martin, J.W., K.A. Crandall, and D.L. Felder (eds.) Decapod Crustacean Phylogenetics. Crustacean Issues. Koenemann, S. (series ed.) Vol. 18. Boca Raton, London, New York: CRC Press, Taylor & Francis Group, 261-279 pp.

Thomas MM (1969a). Notes on some interesting penaeid prawns (Crustacea, Decapoda) from the southeast coast of India. *Journal of the Marine Biological Association of India*, 11(1&2): 191-7.

Thomas MM (1969b). On a new distributional record *of Parapenaeopsis tenella* (Bate) from the south eastern coast of India. *Journal of the Marine Biological Association of India*, 10(1): 166-167.

Thomas MM (1970). *Metapenaeopsis borradaili* (De Man) a penaeid prawn (Decapoda, Penaeidae) new to the Indian Ocean. *Journal of the Marine Biological Association of India*, 12(1&2): 213-216.

Thomas MM (1972). *Trachypenaeopsis minicoyensis* sp. nov. (Penaeidae, Decapoda) from the Laccadive Sea. *Indian Journal of Fisheries*, 19: 116-121.

Thomas MM (1974). Decapod crustaceans new to the Laccadive Archipelago. *Indian Journal of Fisheries*, 21(2): 339-344.

Thomas MM (1976). New records of four Alpheid shrimps from the Indian waters. *Journal of the Marine Biological Association of India,* 18(3): 666-669.

Thomas PA (1986). Prawns of Goa with a note on the biology of *Parapenaeopsis acclivirostris* (Alcock). *Indian Journal of Fisheries*, 33(3): 351-354.

Tirmizi NW (1971). *Marsupenaeus*, a new subgenus of *Penaeus* Fabricius, 1798 (Decapoda, Natantia. *Pakistan Journal of Zoology*, 3: 193-194.

Trivedi DJ, Trivedi JN, Soni GM, Purohit BD and Vachhrajani KD (2015). Crustacean fauna of Gujarat state of India: A review. *Electronic Journal of Environmental Sciences*, 8: 23-31.

Turan C, Gürlek M, Ergüden D, Yağlıoğlu D and Öztürk B (2011). Systematic status of nine mullet species (Mugilidae) in the Mediterranean Sea. *Turkish Journal of Fisheries and Aquatic Sciences*, 11: 315-321.

Unda F (2005). Introduction to Phylogenetics.

Unmesh K and Prakash S (2011). On a documentation of Haddon's carpet anemone (*Stichodactyla haddoni*) (Saville-kent, 1893) (Anthozoa: Actiniaria: Stichodactylidae) and its unique symbiotic fauna from Gulf of Kutch. *Bugs R All. Newsletter of the Invertebrate Conservation and Information Network of South Asia*, 17: 31-34.

Vaitheeswaran T (2014). New Record of *Axiopsis consobrina* (De Man, 1905) (Family: Axiidae:Borradaile, 1903) (Crustacea : Decapoda : Thalassinidea) off Thoothukudi, Southeast Coast of India (08<sup>o</sup> 53.6'N 78<sup>o</sup> 16'E and 08<sup>o</sup> 53.8'N 78<sup>o</sup> 32'E). *Indian Journal of Veterinary and Animal Science Research*, 43 (1): 49-57.

Vaughan DB, Grutter AS and Hutson KS (2018). Cleaner shrimp are a sustainable option to treat parasitic disease in farmed fish. *Scientific reports*, 8(1): 1-10.

Vazquez-Bader AR, Carrero JC, Garcia-Varela M, Gracia A and Laclette JP (2004). Molecular phylogeny of superfamily Penaeoidea Rafinesque-Schmaltz, 1815, based on mitochondrial 16S partial sequence analysis. *Journal of shellfish research*, 23: 911-918. Venkataraman K and Raghunathan C (2015). Coastal and marine biodiversity of India. In Marine faunal diversity in India. Academic Press, 303-348 pp.

Venkataraman K, Jeyabaskaran R, Raghuram KP and Alfred JRB (2004). Bibliography and checklist of corals and coral reef associated organisms of India, *Records of Zoological Survey of India Occasional Paper*, 226:1-468.

Vinagre C, Dias M, Fonseca C, Pinto MT, Cabral HN and Silva A (2015). Use of rocky intertidal pools by shrimp species in a temperate area. *Biologia*, 70(3): 372-379.

Violle C, Navas ML, Vile D, Kazakou E, Fortunel C, Hummel I and Garnier E (2007). Let the concept of trait be functional!. *Oikos*, 116: 882-892.

Voloch CM, Freire PR and Russo CA (2005). Molecular phylogeny of penaeid shrimps inferred from two mitochondrial markers. *Genetics and Molecular Research*, 4(4): 668-674.

Wagner GM, Akwilapo FD, Mrosso S, Ulomi S and Masinde R (2004). Assessment of Marine Biodiversity, Ecosystem Health, and Resource Status in Mangrove Forests in Mnazi Bay Ruvuma Estuary Marine Park; Nairobi, IUCN EARO,106 pp.

Wang YR and Sha ZL (2015). A review of the genus *Synalpheus* (Crustacea: Decapoda: Caridea: Alpheidae) from China seas. *Zoological Systematics*, 40(4): 357-435.

Watson JD and Crick FH (1953). The structure of DNA. *Cold Spring Harbor symposia on Quantitative Biology*, 18: 123-131

Williams AB (1984). Shrimps, lobsters, and crabs of the Atlantic Coast of the Eastern United States, Maine to Florida. Smithsonian Institution Press, Washington, D.C Williams ST, Knowlton N, Weigt LA and Jara JA (2001). Evidence for three major clades within the snapping shrimp genus Alpheus inferred from nuclear and mitochondrial gene sequence data. *Molecular phylogenetics and evolution*, 20(3): 375-389.

Wilson EO (2004). Taxonomy as a fundamental discipline. *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences*, 359(1444): 739.

Wood-Mason J and Alcock A (1891a). Natural history notes from H. M. Indian marine survey steamer" Investigator", Commander R.F. Hoskyn, R.N., commanding. Series II, No. 1. On the results of deep-sea dredging during the season 1890-1891. *The Annals and Magazine of Natural History*, 6(8): 268-286.

Wood-Mason J and Alcock A (1891b). Natural history notes from H. M. Indian marine survey steamer 'Investigator', Commander R.F. Hoskyn, R.N., commanding. No. 21. Note on the results of the last season's deep-sea dredging. *The Annals and Magazine of Natural History*, (7) 6: 186-202.

Wood-Mason J and Alcock A (1891c). Natural history notes from H. M. Indian marine survey steamer" Investigator", Commander R.F. Hoskyn, R.N., commanding. Series II, No. 1. On the results of deep-sea dredging during the season 1890-1891. *The Annals and Magazine of Natural History*, (8)6: 353-362.

Yaldwyn JC and Webber WR (2011). Annotated checklist of New Zealand Decapoda (Arthropoda: Crustacea). *Tuhinga*, 22: 171-272.

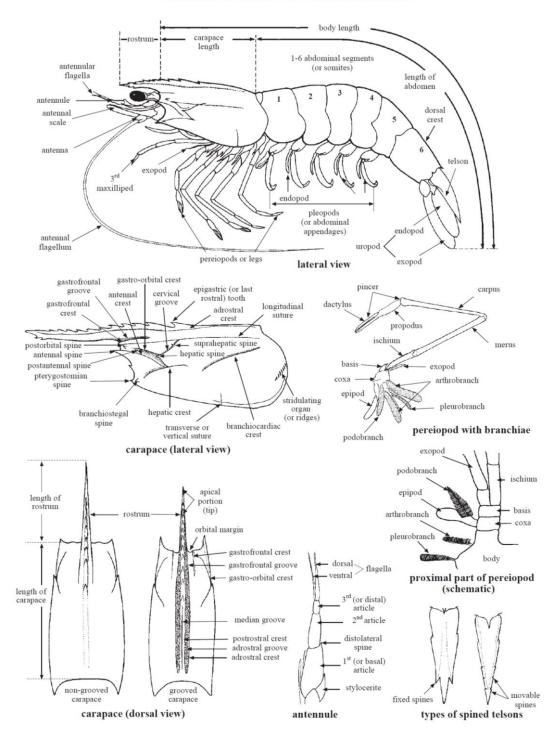
Yang CH, Kumar AB and Chan TY (2017). Further records of the deep-sea pandalid shrimp *Heterocarpus chani* Li, 2006 (Crustacea, Decapoda, Caridea) from southern India. *ZooKeys*, 685: 151-159.

Yang CH, Sha Z, Chan TY and Liu R (2015). Molecular phylogeny of the deep-sea penaeid shrimp genus *Parapenaeus* (Crustacea: Decapoda: Dendrobranchiata). *Zoologica Scripta*, 44(3): 312-323.

Zare P, Ghasemi E and Sarfaraz E (2010). The First Record of *Exopalaemon styliferus* (H. Milne-Edwards, 1840) (Decapoda: Caridea: Palaemonidae) from Iran. *Turkish Journal of Fisheries and Aquatic Sciences*, 10: 523-525.

Zhao Y, Zhu X, Li Y, Han Z, Xu W, Dong J, Wei H and Li X (2019). Mitochondrial genome of Chinese grass shrimp, *Palaemonetes sinensis* and comparison with other Palaemoninae species. *Scientific reports*, 9: 1-10.

# Annexure 1 Technical terms and Measurements use for Prawns and shrimps identification (modified from Holthius, 1980).



TECHNICAL TERMS AND MEASUREMENTS

# Annexure 2: DNeasy Blood and Tissue Kit (Qiagen) Protocol

- The animal was washed before proceeding for DNA isolation using 100% ethanol and then rinsed with sterile distilled water.
- Approximately 20 mg of tissue was dissected and chopped into small pieces. Tissue was transferred into a 1.5 ml sterile microcentrifuge tube, and 180  $\mu$ l ATL buffer was added.
- 20  $\mu$ l of proteinase K was added and mixed thoroughly by vortexing. The samples were incubated at 56°C until the tissue was completely lysed.
- To clear lysate 200 µl buffer AL was added and mixed thoroughly by vortexing than added 200 µl of chilled ethanol (96-100%) to the above solution and mixed again by vortexing.
- A mixture was transferred to the DNeasy minispin column placed into a 2 ml collection tube. Columns centrifuged at 8000 rpm for 1 minute. Flow-through was discarded.
- DNeasyminispin column was placed in a new 2ml collection tube, and 500 µl buffer AW1 was added. Colum was centrifuged at 8000 rpm for 1 minute. Flow-through was discarded.
- DNeasy minispin column was placed in a new 2ml collection tube, and 500 µl buffer AW1 was added. Colum was centrifuged at 8000 rpm for 1 minute. Flow-through was discarded.
- DNeasy minispin column was placed in a new 2ml collection tube, and 500 µl buffer AW2 was added. Colum was centrifuged at 14,000 rpm for 3 minutes to dry the DNeasy membrane. Flow-through and the collection tube were discarded.
- Columns were placed in 1.5 ml microcentrifuge tubes. 35 μl buffer AE was added directly onto the DNeasy membrane and incubated at room temperature (25<sup>o</sup> C) for 5 minutes.
- DNA was eluted by spinning the DNeasy columns at 8000 rpm for 1 minute.
- The step mentioned above was repeated for the yield concentration of DNA. DNA was stored at - 20° C.

## Annexure 3: Agarose gel electrophoresis

## Chemicals for agarose gel electrophoresis

Running buffer- 5X Tris borate EDTA (TBE) buffer

#### Preparation of 5X TBE buffer

Chemical	Amount added
Tris base	54 g
Boric acid	27.5 g
0.5 M EDTA (pH 8)	20 ml

Above mentioned chemicals were mixed in a volumetric flask and the solution was made up to 1 liter by adding distilled water. Buffer was subjected to autoclave treatment, as discussed above. 5X TBE was diluted to 0.5X before to use.

## Preparation of Tris HCl EDTA buffer

Chemical	Required Strength in the	Amount added in 200
	buffer	ml
Tris-HCl	10 mM	0.24g
EDTA	1 mM	0.06g

Two hundred milliliter buffer solution of pH 8.0 was prepared by dissolving above mentioned chemicals.

#### Ethidium bromide (10 mg/l), 10 ml

0.1 g of Ethidium Bromide was added to 10 ml of distilled water and kept on a magnetic stirrer to make sure that the dye has dissolved completely. This was transferred to an amber-colored reagent bottle and stored at 4 °C.

# Plate preparation and casting the gels

The cleaned agarose gel casting cassette and comb were wiped with methanol, and open sides of the tray were sealed with gel sealing tape. The

comb was placed in the given slits of the plate. The calculated amount of agarose in TBE buffer was mixed to prepare a 2 % solution. The agarose was dissolved completely in the buffer by heating the mixture at 80-85°C in the microwave oven and was cooled to 50°C. Ethidium bromide was added in a final concentration of 0.6 mg/ml and mixed well. The liquid was gently poured into the casting tray before it gets solidified. The combs and sealed tape were removed slowly after complete solidification of the agarose gel.

# Qualification and quantification DNA and PCR products

After DNA isolation and PCR amplification, the amplicons were observed on 0.8 and 2% agarose gel. The DNA and PCR product were mixed loading dye and carefully loaded in the wells using gel-loading tips. 100 bp Marker Electrophoresis was carried out at 150V. The gel images were recorded in JPEG or TIF formats using a gel documentation system (Biorad, USA). The gel was examined using the software Image lab version 3.0 (Biorad, USA).