CHAPTER 6

STABILITY STUDIES



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6.1 INTRODUCTION

For liposomal products an attention has been focused on two processes affecting the quality and therefore acceptability of liposomes (Talsma and Crommelin, 1992). Especially, with liposomal product to see the market it should stable during the shelf life (storage or transport). In general, a shelf life of at least one year is a minimum prerequisite criterion for a commercial product First leakage of drug from the vesicles may take place into the extra liposomal compartment. Secondly, there is a possibility of liposomal aggregation and/or fusion, which leads to formation of larger particles (Slabbert et al., 2011; Fang et al., 1997; Cliff et al., 1992; Grit and Crommelin, 1992). These parameters will alter the in vivo fate, affecting therapeutic index of the drug. Hydrolysis of phospholipids is one of the parameters like to cause the formation of fatty acids and lysophopholipids (Grit and Crommelin, 1993; Mowri et al., 1984). Although under dehydrated storage, there is least possibility of the formulation to encounter hydrolytic degradation. Another aspect to be considered is liposome oxidation (Frokjaer et al., 1984). Stability is considered as chemical stability of drug substance in a dosage form. However, the performance of liposomal formulation is not only dependent upon the content of the drug substance, but also dependent on reproducible in vivo performance of the formulations. Formulations under stability studies were considered chemically stable by evaluating the drug leakage from liposomes. The stability protocol was designed as per ICH guidelines (Singh, 1999) for countries falling under zone III (hot, dry) and zone IV (very hot, humid) (US FDA, 2002).

6.2 METHOD

Comparative stability studies were carried out of the potential liposomal formulations at accelerated condition $(25^{\circ}C \pm 2^{\circ}C, 60\% \text{ RH} \pm 5\% \text{ RH})$ for six months and at longterm conditions $(5^{\circ}C \pm 3^{\circ}C)$ up to twelve months. Lyophilized liposomal formulations containing 0.2 mg RGZ and 0.2 mg CDS were filled into amber color glass vials, purged with nitrogen, sealed and stored at the above mentioned condition. At each sampling time different vial is used for the stability testing (Bhalerao and Raje, 2003; Manosroi et al., 2004; Manosroi et al., 2002; Yang et al., 2007; Anderson and Omri, 2004; Changsan et al., 2009; Winterhalter and Lasic, 1993; Ugwu et al., 2005).

Chapter 6 Stability Studies The liposomal formulations were examined visually for the evidence of discoloration. The content of the vial are tested for percentage drug retention (PDR), particle size, zeta-potential, assay and water content. The stability results are summarized in Tables 6.1 (RGZ) and Table 6.2 (CDS).

6.3 RESULTS AND DISCUSSION

The physical stability of liposomes is one of the biggest obstacles in formulation commercially viable product (Fildes, 1981). Liposomes should be stable for 1-2 years preferably at room temperature to be pharmaceutically acceptable with high drug retention within liposome and the particle size should be maintained during storage time, hence the drug leakage, particle size growth, change in zeta-potential and the chemical stability of drugs were studied at accelerated condition $(25^{\circ}C \pm 2^{\circ}C, 60\%$ RH \pm 5% RH) for six months and at long-term conditions (5°C \pm 3°C) up to twelve months. No significant differences were found in all above mentioned parameters at both conditions.

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6.3.1 Stability testing of RGZ liposomal formulation

The stability testing of prepared RGZ liposomal formulation was performed at accelerated condition $(25^{\circ}C \pm 2^{\circ}C, 60\% \text{ RH} \pm 5\% \text{ RH})$ for six months and at long-term conditions $(5^{\circ}C \pm 3^{\circ}C)$ up to twelve months and the effect on various parameters was studied and reported below.

Sampling time (Month)	Description	Assay (%)	Percent drug retained	Water content (%)	Particle size [Z-Average (d.nm)]	Zeta potential (mV)				
Initial	White powder	101.16±2.27	101.55 ± 3.21	3.1 ± 0.21	180.1 ± 2.93	-21.9 ± 1.33				
39938 ⁴⁴ 5757, 20108 2015-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-	Accelerated condition (25°C ± 2°C, 60% RH ± 5% RH)									
1	White powder	101.08 ± 2.45	100.46 ± 3.47	3.2 ± 0.18	183.4 ± 2.74	-21.6 ± 1.27				
2	White powder	101.32 [*] ± 1.98	99.19 ± 2.87	3.2±0.19	186.2 ± 2.22	-20.8 ± 1.43				
3	White powder	100.73 ± 3.03	98.77 ± 2.63	3.3 ± 0.21	188.7 ± 3.19	-20.9 ± 1.51				
6	White powder	99.79 ± 3.28	97.66 ± 3.07	3.4 ± 0.24	194.3 ± 3.35	-19.7 ± 1.20				
Long-term conditions (5°C ± 3°C)										
1	White powder	101.77 ± 2.83	101.81 ± 2.26	3.1 ± 0.19	180.4 ± 3.02	-22.1 ± 1.21				
2	White powder	101.43 ± 2.42	101.07 ± 3.09	3.1 ± 0.22	179.2 ± 3.11	-21.8 ± 1.49				
. 3	White powder	101.09 ± 3.11	101.24 ± 3.13	3.1 ± 0.22	183.3 ± 2.95	21.3 ± 1.46				
6	White powder	101.82 ± 2.07	100.92 ± 3.42	3.2 ± 0.28	182.4 ± 2.84	-21.1 ± 1.13				
9	White powder	100.68 ± 1.84	100.76 ± 2.86	3.2 ± 0.29	184.6 ± 2.50	-21.3 ± 1.59				
12	White powder	100.31 ± 3.21	100.01 ± 2.29	3,2 ± 0.24	186.2 ± 3.14	-20.7 ± 1.09				

Table 6.1 Stability testing data of RGZ liposomal formulation

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6.3.2 Stability testing of CDS liposomal formulation

The stability testing of prepared CDS liposomal formulation was performed at accelerated condition $(25^{\circ}C \pm 2^{\circ}C, 60\% \text{ RH} \pm 5\% \text{ RH})$ for six months and at long-term conditions $(5^{\circ}C \pm 3^{\circ}C)$ up to twelve months and the effect on various parameters was studied and reported below.

Sampling time (Month)	Description	Assay (%)	Percent drug	Water content (%)	Particle size [Z-Average (d:nm)]	Zeta potential (mV)				
Initial .	White powder	99.82 ± 2.53	98.93 ± 3.38	2.7 ± 0.19	184.6 ± 3.01	-29.3 ± 1.41				
		Accelerated condi	tion (25°C ± 2°C, 6	0% RH ± 5%	RH)					
1 `	White powder	99.98 ± 2.21	98.12 ± 3.19	2.7 ± 0.23	185.1 ± 2.69	-28.5 ± 1.38				
2	White powder	99.76 ± 2.33	96.74 ± 3.26	2.7 ± 0.23	188.4 ± 2.09	-27.6 ± 1.26				
3	White powder	99.41 ± 2.72	96.39 ± 2.87	2.8±0.19	190.3 ± 2.84	-27.3 ± 2.01				
6	White powder	98.63 ± 1.97	95.28 ± 2.82	2.9±0.20	196.8 ± 3.09	-26.2 ± 1.42				
	Long-term conditions (5°C ± 3°C)									
1	White powder	99.54 ± 3.02	98.47 ± 2.71	2.7±0.18	183.9 ± 3.25	-29.2 ± 1.19				
- 2	White powder	99.61 ± 2.21	98.59 ± 3.31	2.7 ± 0.25	184.7 ± 2.88	-29.6 ± 1.44				
3 ·	White powder	99.71 ± 2.95	98.38 ± 3.47	2.7 ± 0.23	185.5 ± 2.61	-28.1 ± 1.37				
6	White powder	99.34 ± 3.16	98.16 ± 2.56	2.7 ± 0.31	187.2 ± 3.37	-27.6 ± 1.33				
9	White powder	99.28 ± 2.78	97.87 ± 2.72	2.7 ± 0.25	189.9 ± 3.00	-27.4 ± 1.72				
12	White powder	98.94 ± 2.45	97.12 ± 3.01	2.8±0.27	193.1 ± 2.98	-26.9 ± 1.21				

Table 6.2 Stability testing data of CDS liposomal formulation

6.4 CONCLUSION

The decrease in drug assay, percentage drug retained, and zeta-potential and increase in water content, particle size were observed at accelerated condition $(25^{\circ}C \pm 2^{\circ}C, 60\% \text{ RH} \pm 5\% \text{ RH})$ for six months and at long-term conditions $(5^{\circ}C \pm 3^{\circ}C)$ up to twelve months for both RGZ and CDS liposomal formulation but the changes were statistically insignificant. Hence, both the formulations were considered as stable and were selected for further studies.

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6.5 REFERENCES

Anderson M, Omri A. The effect of different lipid components on the in vitro stability and release kinetics of liposome formulations. Drug Deliv. 2004 Jan-Feb;11(1):33-9.

Bhalerao SS, Raje Harshal A. Preparation, optimization, characterization, and stability studies of salicylic acid liposomes. Drug Dev Ind Pharm. 2003 Apr;29(4):451-67.

Changsan N, Chan HK, Separovic F, Srichana T. Physicochemical characterization and stability of rifampicin liposome dry powder formulations for inhalation. J Pharm Sci. 2009 Feb;98(2):628-39.

Cliff RO, Ligler F, Goins B, Hoffmann PM, Spielberg H, Rudolph AS. Liposome encapsulated hemoglobin: long-term storage stability and in vivo characterization. Biomater Artif Cells Immobilization Biotechnol. 1992;20(2-4):619-26.

Fang JY, Lin HH, Hsu LR, Tsai YH. Characterization and stability of various liposome-encapsulated enoxacin formulations. Chem Pharm Bull (Tokyo). 1997 Sep;45(9):1504-9.

Fildes FJT (1981) Liposomes: The Industrial view point. In: Liposomes from Physical Structure to Therapeutic Applications, Knight CG (Ed), Elsevier Biomedical Press, New York, 465-483.

Frokjaer S, Hjorth EL, Worts O. (1984) Stability testing of liposomes during storage. In: Liposome Technology: Preparation of liposomes, Gregoriadis G (Ed), CRC Press, Boca Raton, Florida, 235-245.

Grit M, Crommelin DJ. The effect of aging on the physical stability of liposome dispersions. Chem Phys Lipids. 1992 Sep;62(2):113-22.

Grit M, Zuidam NJ, Underberg WJ, Crommelin DJ. Hydrolysis of partially saturated egg phosphatidylcholine in aqueous liposome dispersions and the effect of cholesterol incorporation on hydrolysis kinetics. J Pharm Pharmacol. 1993 Jun;45(6):490-5.

Manosroi A, Kongkaneramit L, Manosroi J. Characterization of amphotericin B liposome formulations. Drug Dev Ind Pharm. 2004 May;30(5):535-43.

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Manośroj A, Pedjanasoonthon K, Manosroi J. Stability and release of topical tranexamic acid liposome formulations. J Cosmet Sci. 2002 Nov-Dec;53(6):375-86.

Mowri H, Nojima S, Inoue K. Effect of lipid composition of liposomes on their sensitivity to peroxidation. J Biochem. 1984 Feb;95(2):551-8.

Singh S. Drug stability testing and shelf-life determination according to international guidelines. Pharm. Technol. 1999;23:68-88.

Slabbert C, Plessis LH, Kotze AF. Evaluation of the physical properties and stability of two lipid drug delivery systems containing mefloquine. Int J Pharm. 2011 May 16;409(1-2):209-15.

Talsma H, Crommelin DJA. Liposomes as Drug Delivery Systems, Part III: Stabilization. Pharm. Technol. 1992;17:48-59.

Ugwu S, Zhang A, Parmar M, Miller B, Sardone T, Peikov V, Ahmad I. Preparation, characterization, and stability of liposome-based formulations of mitoxantrone. Drug Dev Ind Pharm. 2005 Jan;31(2):223-9.

US FDA (CDER), (2002), Draft Guidance for Industry Liposome Drug Products. CMC Documentation.

Winterhalter M, Lasic DD. Liposome stability and formation: experimental parameters and theories on the size distribution. Chem Phys Lipids. 1993 Sep;64(1-3):35-43.

Yang T, Cui FD, Choi MK, Lin H, Chung SJ, Shim CK, Kim DD. Liposome formulation of paclitaxel with enhanced solubility and stability. Drug Deliv. 2007 Jul;14(5):301-8.