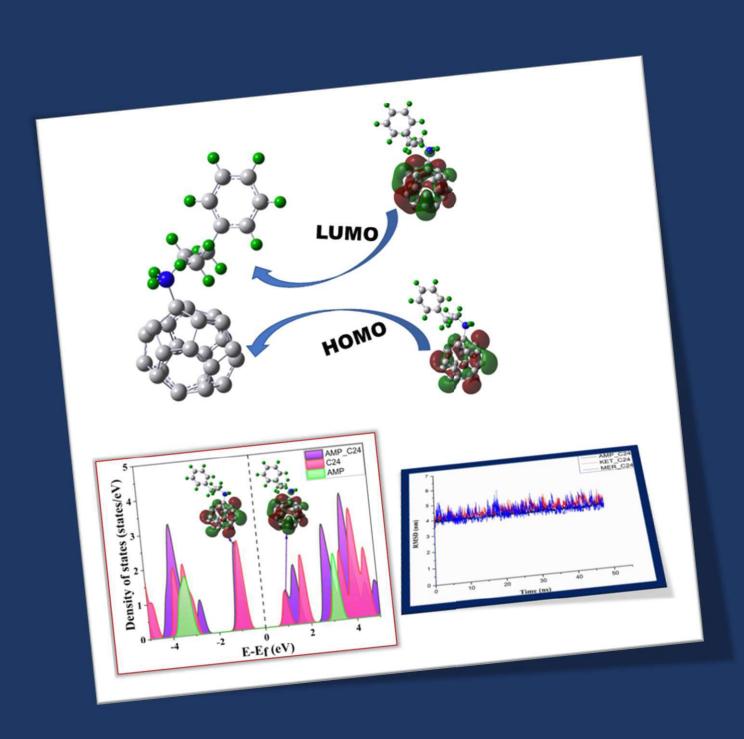
Chapter 4

Exploring C₂₄ Fullerene as an illicit Drug sensor



4.1 Introduction

Concern over tainted pharmaceutical goods is on the rise, and there is now a global issue and conundrum around the pharmaceutical industry's dubious clinical obligations. An accurate evaluation of a medicine's active component content determines the quality of the drug[1]. Identifying and handling illicit drugs, particularly synthetic ones, is a significant and pressing issue in today's world. This challenge poses a threat to social unity, family stability, and human well-being[2-3]. The overdose of some of the drugs can lead to fatal human life, the renowned abuse drugs such as Amphetamine (AMP), Ketamine (KET) and Mercaptopurine (MER). It is very important for drug communities and security agencies alike to detect the drug molecule AMP in biological samples as it is a popular drug that is misused and is used in many ways. Excessive consumption of it leads to various critical issues, including psychological disturbances and an increased risk of traffic accidents. [4]. There is a second fatal drug molecule, KET, which enkindles NMDA receptors on neuronal cells, hence causing hallucinations[5]. Due to the hallucinogenic effects of its excessive use, a K-hole occurs in the human body, resulting in a cataleptic state [6-7]. MER, the last drug from the above group, exhibits immunosuppressive properties and is used for treating acute lymphoblastic leukaemia[8]. Pregnant women are prohibited from taking MER, as described in the previous report. This drug causes abnormal abortion rates during the first trimester of pregnancy [9]. Its cytotoxic nature also limits its application due to several side effects[10]. It's hard to detect these spike drugs because they are queered into quenchers to incapacitate victims, as well as because of their low concentrations and complex sample matrices. For the medical field and for the prevention and reduction of drug-related crimes, it is of great importance to increase research, impressionable, rapid, and inexpensive methods for the detection of illicit drugs[11]. The detection of the drugs has been accomplished using a wide variety of techniques, including high-performance chromatography liquid (HPLC)[12],GCM[13], capillary electrophoresis[14], ion mobility spectrometry[15], UV-visible spectrophotometry[16], electrochemical luminescence[17], Raman spectrometry[18], fluorescence[19], and film-based fluorescent[20]. Despite their time-consuming and expensive instrumentation, most of the techniques mentioned above have been confirmed to rigorous use[11]. Since electrochemical sensors offer a number of advantages, including cost-effectiveness, portability, simplicity, speed, and sensitivity[21], they are an excellent alternative to conventional sensors. A stateof-the-art density functional theory (DFT) approach has been used to detect unscriptural drugs using nanomaterials of low dimension. AMP drug adsorption behavior over pristine and doped Al₁₂N₁₂ and Al₁₂P₁₂ nanocages[22], intrinsic and extrinsic C₆₀ fullerenes[23], Al-doped ZnO nanotubes[24], silicon carbide nanotubes[25], AlN nano-cage[26], and BC₃ nanotubes and nanosheets[27] has been studied by theoretical studies. A number of different materials have been used to investigate the sensing mechanism of MER drug, such as pristine and doped C₇₀ fullerenes[28], boron nitride nanocages[29], BN nanotubes, nanosheets, and clusters[30], as well as B₁₂N₁₂, AlB₁₁N₁₂ and GaB₁₁N₁₂ nanoclusters[31]. R. Zhiani and colleagues undertook a joint study employing both Density Functional Theory (DFT) and Molecular Dynamics (MD) simulations to investigate KET drug detection using functionalized single-walled carbon nanotubes[32]. As compared to other carbon-based nanomaterials like graphene and CNTs, fullerenes have recently received a great deal of attention due to their low side effects in biological media, their high surface area and hydrophobic nature[33-34]. The C₂₄ in Comparison to C₆₀, C₂₀, and doped C₂₀ demonstrates the better sensitivity due to its lower energy band gap[23][35]. As we can see from our literature review, the stability of C_{24} fullerene has proved to be an important factor to ensure its success as a drug carrier and sensor. To the best of our knowledge no systematic study has been conducted on the adsorption of Amphetamine (AMP), Ketamine (KET) and Mercaptopurine (MER) over carbon nanostructures using MD simulations. Using classical MD simulations and DFT theories, we have investigated sensing applications of the pristine C₂₄ fullerene for the above-mentioned drug molecules. In order to determine whether the pristine C₂₄ fullerene is suitable for sensing, we conducted DFT as well as classical MD simulations on the above-mentioned drug molecules. Additionally, a DFT investigation was conducted to assess the binding mechanisms and sensitivity of three drug molecules when interacting with C24 fullerene, both in a gaseous environment and under the influence of solvent effects. A DFT study was also used to analyse electronic features such as NBO, Mulliken charge, and DOS. Classical molecular dynamics (MD) simulations were employed to explore the stability and the formation or breaking of bonds within complexes of drug molecules and fullerene at a temperature of 310K (room temperature).

4.2 Computational Details:

4.2.1 Details of the DFT calculation

The structural refinement of both the drug molecules and C₂₄ fullerene, along with electronic computations, were conducted using the Gaussian 09 software package[36] were performed using the DFT/B3LYP technique employing the 6-31G(d,p) basis set together with the D3 version of Grimme's dispersion[37]. The adsorption energy, E_{ad} is defined as

$$E_{ad} = E_{C24/Drug\ Molecules} - (E_{C24} + E_{Drug\ Molecules}).....(1)$$

Here, $E_{Drug\ Molecules}$ and $E_{C24/Drug\ Molecules}$ are the energies of the isolated drug molecules and optimized C_{24} fullerene with the adsorbed drug molecules, respectively. E_{C24} corresponds to the energy of the optimized C_{24} fullerene. The difference between the lowest unoccupied molecular orbital (LUMO) and highest molecular orbital (HOMO), which is known as the HOMO-LUMO energy gap E_G , is written as

$$E_G = E_{LUMO} - E_{HOMO}$$
 (2)

4.2.2 Details of the MD Simulation

We employed a classical molecular dynamics simulation, utilizing the GROMACS package version 5.0.2[38] and the CHARRM27 force field [39], to validate the adsorption behaviour of drug molecules Amphetamine (AMP), Ketamine (KET), and Mercaptopurine (MER) on the external surface of C₂₄ fullerene. Three simulation systems are developed (C₂₄/AMP, C₂₄/KET, and C₂₄/MER). Table 4.6 lists details of all investigated systems. It is shown that the TIP3P model can be used to describe the structure of water molecules [40]. For the MD simulation, we employed the ultimate optimized structures derived from our DFT computations. These structures were then converted into PDB format via GaussView 5. To account for periodic boundary conditions, we ensured that all simulation boxes were sufficiently large, thereby mitigating interactions between components and neighbouring cells. It's important to highlight that in each of the containers, the drug molecules were positioned approximately 1.5-2 Å apart from the surfaces of the C24 fullerenes. Prior to equilibrating the systems using NVT and NPT ensembles for 100 picoseconds, a preliminary minimization stage was performed on the entire system. By using V-rescale and Berendsen algorithms, the pressure and temperature are kept constant at 310 K and 1 bar respectively [41]. Production MD runs with a step time of 2fs have been carried out for each system for 50ns under periodic conditions. The LINCS method has

been employed[42] to constrain all bonds to their equilibrium length. Long-range electrostatic interactions have been treated using the particle-mesh Ewald (PME) approach, and non-bonded interactions are estimated with a 1.2 nm range cut-off [43]. The molecular visualization is done using the Visual Molecular Dynamics (VMD) program[44].

4.3 Results and Discussion

4.3.1 DFT calculation results

4.3.1.1 Geometry Optimization of the adsorbent C24 fullerene and Drug molecules

This section seeks to provide a thorough explanation of how the three chosen drug molecules interact with C₂₄ fullerene. Using DFT-D3/B3LYP,6-31G(d,p), all of the chosen molecular structures, including C₂₄ fullerene and all three drug molecules, were relaxed in the first step. The optimized geometries are depicted in Figure 4.1. The C₂₄ fullerene with a D_{6h} point group has one hexagonal ring at the top and one at the bottom, and twelve pentagonal rings in the centre. The optimal bond lengths for C=C and C-C are 1.36 Å and 1.52 Å, while Energy gap (E_G) is 1.82 eV with HOMO and LUMO energies of -5.65 and -3.83 eV, respectively (see Table 4.1). The optimized bond lengths and energy gap are largely in line with the available data[45]. The calculation of the formation energy, which was determined to be -8.29 eV/atom, revealed the structural stability of C₂₄, which is consistent with existing literature [46-47]. The formation energy is highly negative, which ensures stability. In addition, vibrational modes have been calculated to confirm stability with infrared frequencies, and there is no imaginary frequency present. Vibrational frequencies vary between 200 and 1600 cm⁻¹, which is consistent with Hossian et.al. [46]. We optimized the drug molecules after confirming the basic properties of C₂₄. It is necessary to have a precise representation of the charge population of the drug molecules in order to better understand their binding affinity and interaction with C₂₄. As such, Figure 4.2 illustrates the calculated Molecular Electrostatic Potential (MESP) for the aforementioned drug molecules and fullerene C₂₄. A colour scheme has been chosen in such a way that the red part represents attractive potential and the blue part represents repulsive potential. C24 fullerenes contain intermediate negative portions over both hollow pentagonal and hexagonal rings, making them ideal candidates for nucleophilic attack sites. Due to the lone pair electrons of nitrogen atoms, a high negative electron density surface is visible along the nitrogen atom's NH₂ group and an intermediate negative region along the other group of AMP drug. In addition, KET and MER molecules contain regions with high

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electron density along oxygen and sulphur atoms. The structures of C_{24} , AMP, KET, and MER molecules do not change significantly when optimized in water.

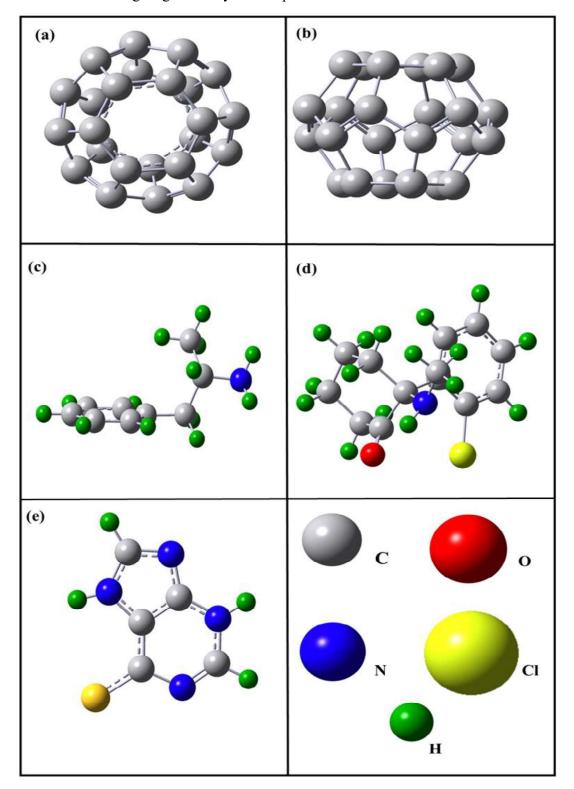


Figure 4.1: Optimized geometry of C_{24} fullerene (top(a) and side view (b)) and drug molecules (AMP (c), KET (d),) and MER (e)).

4.3.1.2 Interaction characteristics of Amphetamine (AMP), Ketamine (KET) and Mercaptopurine (MER) drug molecules onto C₂₄ fullerene

In this study, we analysed the most stable configuration of drug molecules interacting with C_{24} fullerene to verify their affinity towards C_{24} fullerene (**Figure 4.3**). A study shows that the drug molecules are adsorbable over hexagonal or pentagonal rings of the C_{24} fullerene without any structural distortion. A number of possible adsorption sites were explored for the drug. Several possible adsorption sites have been proposed for the drug molecules on C_{24} in order to find the minimum energetic conformer. As a result of the additional electrons present,

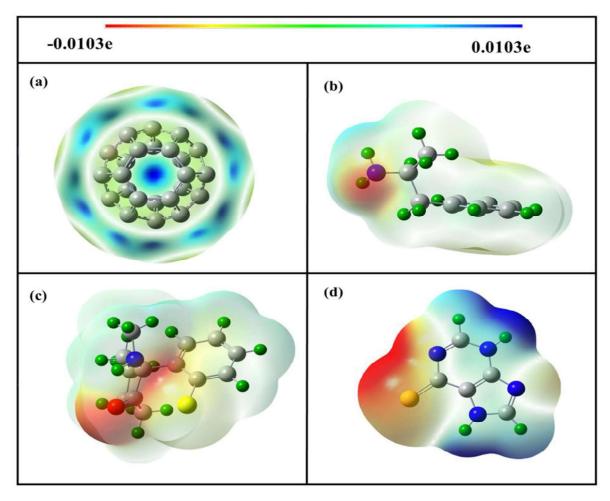


Figure 4.2: Electrostatic potential (ESP) maps for C_{24} fullerene (a) and drug molecules AMP (b), KET (c), and MER (d) (EPS values ranging from -0.0103e to 0.0103e).

N-atoms are preferred over H-atoms for drug adsorption onto C_{24} , and consistent with the results of MESP analysis. The distance between the nitrogen (N) atom of AMP and the carbon (C) atom of C_{24} is roughly 1.55 Å, and the drug molecule AMP's adsorption energy onto C_{24} is

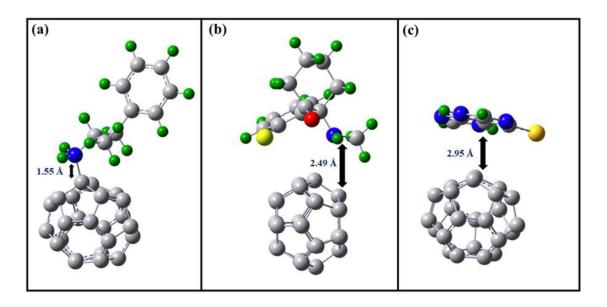


Figure 4.3 : The lowest-energy configurations of AMP (a), KET (b), and MER (c) when adsorbed onto C₂₄ fullerene, respectively.

-1.02 eV (see **Table 4.1**). Chemisorption is the process by which the AMP molecule is strongly adsorbed on the C_{24} fullerene[48]. With -0.423 eV energy, KET drug molecule adsorbed over the pentagonal ring of C_{24} and optimized separation between **KET** drug molecule and C_{24} fullerene is 2.49 Å, which indicates its physisorption mechanism. Adsorption of the MER drug molecule occurs through physisorption over the hexagonal site of fullerene C_{24} [35]. The MER drug molecule and C_{24} fullerene are separated by a minimum of 2.95 Å atom to atom. The adsorption energies reveal its nature and the sequence or tendency of drug molecules towards the fullerene.

Table 4.1: Calculated value of Adsorption energy (E_{ad}) , LUMO energy (E_{LUMO}) , Fermi level (E_F) , HOMO energy (E_{HOMO}) , HOMO-LUMO gap (E_G) , relative change in E_F (ΔE_{FR}), relative change in E_G (ΔE_{GR}) and adsorption distance(d) of AMP, KET and MER adsorbed over C_{24} fullerene, respectively.

System	E _{ad} (eV)	ELUMO (eV)	E _F (eV)	Еномо (eV)	E _G (eV)	ΔE _{FR} (%)	ΔE _{GR} (%)	d (Å)
C24		-3.83	-4.74	-5.65	1.82			
AMP/C ₂₄	-1.02	-2.47	-3.42	-4.37	1.90	-27.85	4.39	1.55
KET/C24	-0.423	-3.23	-4.13	-5.02	1.79	-12.87	-1.65	2.49
MER/C ₂₄	-0.365	-3.69	-4.58	-5.46	1.77	-3.37	-2.74	2.95

4.3.1.3 The natural bond orbital (NBO) charge analysis

The ability to better comprehend how adsorbate and adsorbent interact is made possible by the electronic charge transfer, which is a crucial component. The NBO analysis, which is displayed in **Table 4.2**, can be used to explain the charge transfer between drug molecules and C_{24} in their ideal geometrics throughout the adsorption process[49]. We also examined the NBO(Q_N)[50] and Mulliken charge transfer (Q_M)[50-51] between the drug molecules and C_{24} fullerene to ascertain the path of charge transfer within the molecule-molecule interactions. Our results from the Mulliken charge analysis (Q_M) are consistent with the positive value of Q_N that we obtain. These were also noted in earlier literature[51], and it is believed that charge was transferred from drug molecules to C_{24} fullerene. The charge distribution of drug molecules adsorbed over C_{24} is shown in **Figure 4.4**. Additionally, the NBO calculation has been used to clarify the second-order perturbation stabilization energy, $E^{(2)}$, of the donoracceptor (bond-antibond) interaction between the C_{24} fullerene and drug molecules, which was predicted by the second-order Fock matrix[49]. For the delocalization of the donor (i) to the acceptor (j), the equation of second order perturbation stabilization energy, $E^{(2)}$, is represented as

The second order perturbation stabilization energy is directly correlated with the NBO interacting intensity, and it is well known that a stronger interaction between the donor and acceptor results in a higher stabilization energy, which confers more stability for C_{24} /drugmolecule complexes. The greatest $E^{(2)}$ values are shown in **Table 4.2**. According to the obtained

Table 4.2: The NBO second-order perturbation energy $(E^2, kcal/mol)$ corresponds to the charge transfer between the C_{24} fullerene and AMP, KET, and MER, respectively.

System	Donor	Acceptor	E ² (Kcal/mol)	
AMP/C ₂₄	BD(C3-C6)	BD*(N25-C26)	46.40	
KET/C ₂₄	LP(N27)	BD*(C7-C15)	9.33	
MER/C ₂₄	BD*(C30-C31)	BD*(C9-C10)	2.60	

results, the intermolecular interaction in the AMP/C₂₄ complex identifies the interaction as (C3-C6) of C₂₄ fullerene functions as donor while (N25-C26) of AMP acts as acceptor, which is compatible with the results of ESP. The highest value of $E^{(2)}$ is 46.40 kcal/mol, indicating that there is strongest interaction between the AMP drug and the C₂₄ fullerene. Chemisorption is taking place which confirms from the charge transfer and adsorption energy of AMP over C₂₄ fullerene. The greatest values of $E^{(2)}$ for the complexes C₂₄/KET and C₂₄/MER are 9.33 and 2.60 Kcal/mol, respectively. The lower values in comparison to the AMP/C₂₄ combination support the physisorption nature of their adsorption. In general, it may be said that most interactions take place between the donor orbitals of the drug molecule and the acceptor orbitals of pristine C₂₄.

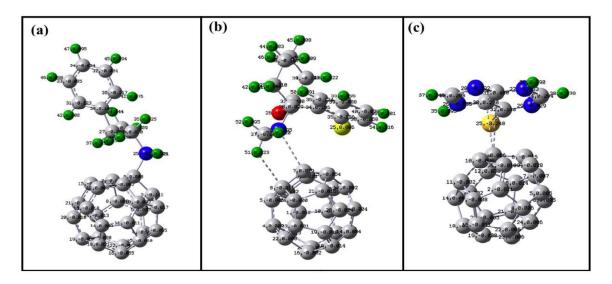


Figure 4.4: Mulliken charge distribution analysis for AMP (a), KET (b), and MER (e) when adsorbed onto C_{24} fullerene, respectively.

4.3.1.4 Density of states (DOS)

The change in the HOMO and LUMO levels, which causes the charge transfer process, provides evidence of the drug-system interaction. As a result, we generated the density of states using GaussSum software and calculated the energy gap of the systems using equation 2. To facilitate a comparison between the HOMO and LUMO levels of the pristine C_{24} and their corresponding states when interacting with unaltered drugs, we computed the Fermi level. This was achieved by calculating half the sum of the HOMO and LUMO levels within the systems and setting it as the zero-reference point on the density of states plot. This enabled us to assess the changes in the HOMO and LUMO levels of pristine C_{24} following their interaction. Two

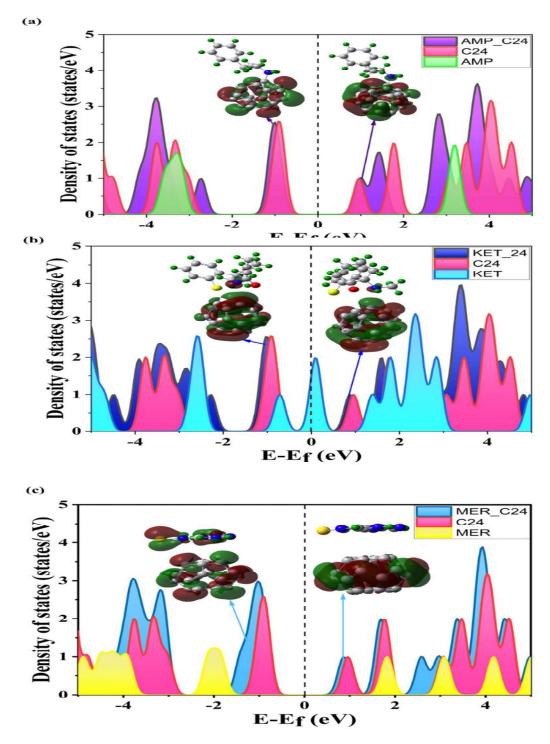


Figure 4.5: Density of states for the interaction between C_{24} fullerene and AMP (a), KET (b), and MER (c).

other parameters that affect the energy gap are the system stability and chemical reactivity. Higher energy gap corresponds to high kinetic stability, which in turn denotes low chemical reactivity, and vice versa. The confirmed energy gap of C₂₄ is 1.82 eV, which is consistent with

our earlier research[35] and with previously reported works[45]. The energy gaps for the drug molecules AMP, KET, and MER are 6.24 eV, 5.18 eV, and 3.69 eV, respectively. Higher energy gaps suggest that electrons from the low-lying HOMO must be transferred to the high-lying LUMO, which is energetically unfavourable and indicates that C₂₄ is highly reactive to these drugs. It's noteworthy to note that after analysing the density of states of C24 interacting with these three drugs, we found that the energy gap of the complex that was created after C₂₄ interacted with AMP showed an increase. Based on the shift of the HOMO and LUMO levels to negative values, the energy bands of these systems increase compared to the C₂₄ molecules (Figure 4.5). We notice the greatest change in HOMO level in the AMP instance, as well as the localization of both HOMO and LUMO at C24 fullerene. This change in HOMO and LUMO and the Mulliken charge analysis conclude that there is charge transfer from C24 fullerene to the AMP drug molecule. In comparison to the KET and MER complex configurations, the E_G corresponds to the AMP complex configuration having the highest value. However, we find that the MER complex configuration has the smallest energy gap compared to the other two complex configurations due to the lowest negative values for HOMO and LUMO (see Table-1). In the case of the MER and KET complexes, we also notice that the HOMO and LUMO energy levels are not significantly changed, and there is a density of states that satisfy the physisorption nature. However, HOMO and LUMO localize At the C₂₄ fullerene in every instance. As MER molecules have a lower HOMO level than AMP molecules—nearer to C₂₄ molecules—the charge transfer mechanism is considerably simpler with MER than it is with AMP. The interaction process between the complexes of the considered minimal configurations is confirmed by the alternation of the density of the states of the complexes. According to Figure 4.5(a), the major contribution to the Fermi level HOMO and LUMO levels comes from C₂₄ fullerene, which shows that AMP is minimally hybridized with C₂₄ fullerene in the AMP-C₂₄ complex[52]. Figure 5(b) makes this clear that the ketamine molecule's density of state is close to the Fermi level from DOS and molecular orbit level, and after the interaction disappears, there is a shift of the LUMO side close to the Fermi level, which is caused by the hybridization of ketamine and C_{24} fullerene. The HOMO near the Fermi level in the AMP C_{24} , on the other hand, is primarily contributed by C₂₄ fullerene, whereas the LUMO near the Fermi level is contributed by C24 and S-N and 6-ring benzene, which is a minimal indication of hybridization with the appearance of the shoulder in the DOS of MER C_{24} (see Figure 5(c)). In Figure 4.6, the localization of the LUMO electron density is shown between the nitrogen

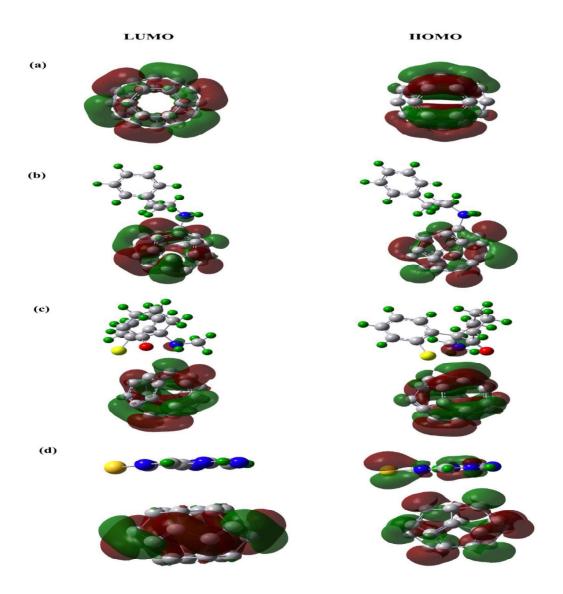


Figure 4.6: The electron density of the Highest Occupied Molecular Orbital (HOMO) and the Lowest Unoccupied Molecular Orbital (LUMO) for the pristine C_{24} fullerene (depicted in (a)), as well as for AMP (b), KET (c), and MER (d) when adsorbed onto C_{24} fullerenes, respectively.

atom of the drug's AMP molecule and the closest carbon atom of the C_{24} fullerene. This occurrence shows that AMP and C_{24} fullerene interact strongly, supporting the existence of a chemical bond. HOMO and LUMO electron densities localize for the KET and MER adsorbed over C_{24} fullerene, suggesting the optimal adsorption energy between them. The charge transfer process within molecules and between two interacting molecules can be understood with the help of the HOMO and LUMO orbitals. HOMO and LUMO represent the electron-donating and electron-accepting capabilities, respectively. It is clear from the C_{24} /AMP complexes that the Nitrogen atoms of the NH₂ functional group of AMP contribute to the LUMO orbital level.

While in the instance of the MER/C₂₄ combination, the HOMO contribution from the S atom and 6-ring benzene shows its contribution in both the HOMO and LUMO orbital levels, a similar NH₂ functional group does not.

4.3.1.5 Sensing Response

The nobleness of a chemical sensor, which related to the energy gap (E_G), can be pursued with great understanding using experimental sensing properties[53]. On the basis of the following equation, the E_G can investigate changes in the electrical conductivity or resistivity of C_{24} fullerene caused by changes in its electronic properties[54-55].

$$\sigma = AT^{3/2} \exp(-E_G/KT)$$
....(4)

Where A is the constant, K is the Boltzmann constant, T is the operating temperature, and σ is the electrical conductivity. Here, we calculate the sensing response using equation (4), which uses the below formula[54] to demonstrate the effectiveness of the sensing materials.

$$S = |(\sigma_1/\sigma_2)-1| = \exp(|\Delta E_G|/KT)-1....(5)$$

Where $\Delta E_G = E^{G2} - E^{G1}$, E^{G2} and E^{G1} defines the energy gaps of the C_{24} nanocage and the complex, respectively.

Table 4.3: Computed variations in E_G (ΔE_G), recovery time (τ), and sensing response (S) for Adenine, Cytosine, Guanine, Thymine, and Uracil when they are adsorbed onto C_{24} fullerene, respectively.

System	ΔE _G (eV)	τ (sec)	S
AMP/C ₂₄	-0.08	38.29	18.98
KET/C ₂₄	0.03	7.54×10^{-9}	2.07
MER/C ₂₄	0.05	8.59×10^{-10}	5.49

AMP considerably changed the HOMO-LUMO band gap of the C_{24} fullerene (Table 4). This causes the E_G of the fullerene to alter significantly as a result of the drug AMP adsorption, revealing that the C_{24} has considerable sensitivity to the drug AMP. C_{24} fullerene is found to have the best sensing response to AMP based on Eq (5) at 310 K, among all the considered

drugs (**Table 4.3**). As a result of not a significant change in E_G after KET and MER adsorb over C_{24} , the sensing performance of those molecules is poor.

4.3.1.6 Recovery Time

Experimentally, recovery time is an essential characteristic for a chemical sensor that reflects the desorption process by heating the adsorbent to a higher temperature or by exposing it to UV radiation and is strongly related to interaction intensity. Higher interaction energies between molecules and adsorbent reduce the recovery time and decrease their own interactions, which makes them less ideal for use in sensing applications. As a result, a good sensor should have a suitable optimal interaction energy and interaction distance [48].

The following equation [56] yields the Recovery time (τ) .

$$\tau = v_0^{-1} \exp(-E_{ad}/KT)....(6)$$

Where T, K, and v_0 , respectively, stand for temperature, Boltzmann's constant, and attempt frequency. Experimental evidence has shown that various photonic frequencies (v_0) or thermal energy have been employed to cause the desorption of drug and biomolecules[56]. By selecting a v_0 of 10^{15} s⁻¹ and a temperature of 300 K, we projected the recovery durations for the AMP, KET, and MER from the C_{24} perimeter (See **Table 4.3**). The highest recovery time for the AMP drug molecule is 38.29 s, which is in accordance with significant chemisorption nature. Due to the longer Recovery time, it is possible to isolate the drug molecule AMP from the sequence of other drug molecules using the C_{24} fullerene. Even for KET and MER, the interatomic distance from the C_{24} fullerene is 2.49 Å and 2.95 Å, and the recovery times are 7.54×10^{-9} s and 8.59×10^{-10} s, respectively. This suggests that the medications KET and MER may be used for the best detection using the C_{24} fullerene.

4.3.1.7 Solvent effect:

The polarizable continuum model, or PCM approach, has been applied in the current investigation to assess the solvent's dominance since the drug/C₂₄ fullerene complexes truly exist biologically[57]. We reoptimized the three drug molecules, C₂₄ individually, and all the minimum configurations of AMP, KET, MER, and C₂₄ complexes in the water solvent using the DFT/B3LYP/6-31(d,p) with D3 version of Grimme's dispersion. The measured adsorption energies and sensor parameter values are listed in Table 4. The results demonstrate that the adsorption energies of the AMP/C₂₄ and KET/C₂₄ complexes relative

to the gaseous phase, which are 1.53 eV and 1.29 eV, respectively, drop when the overall energy values of the C_{24} fullerene, AMP, and KET systems decrease. This is because the solvent has a larger electric permittivity[58]. Additionally, the solvent's impact causes the adsorption energy of MER/ C_{24} complexes to rise, which decreases sensing response and decreases recovery time in an aquatic environment. It is predicted that AMP/ C_{24} and KET/ C_{24} complexes will perform better in an aqueous environment, since lower interaction energies will result in stronger sensing responses and quicker recovery times (see **Table 4.4**). The following equation has been used to compute the solvation energies (ΔE_{sol}) in order to predict the solubility of the drug molecules in water[59].

$$\Delta E_{sol} = E_{sol} - E_{gas} \dots (7)$$

 E_{sol} and E_{gas} are the system's energy in the water solvent and gas, respectively. The measured E_{sol} for the C_{24} fullerene is -0.017 eV, while it is -0.16 eV and -0.24 eV for the AMP and KET, respectively. ΔE_{sol} observed for AMP/ C_{24} and KET/ C_{24} complex is -0.69 eV and -1.13 eV respectively. AMP and KET complexes with C_{24} have solvation energies that are less negative than the total solvation energies of AMP, KET, and C_{24} fullerene. C_{24} fullerene, AMP, and KET drug molecules appear to be highly polar and solubilized in water. On the basis of the electronic structure, the E_G change for C_{24} fullerene by AMP, KET and MER increases in water, 0.16,0.26 and 0.03 eV respectively, when compared to gas. The C_{24} fullerene is highly to solvent medium as its sensing response has increased from 18.98 and 2.07 in the gas phase to 7310.04 and 4964.69 in water respectively. Sensing response, however, decreased in the case of the MER drug (2.19), indicating that it is less interactive in solvent media. Response (S) for AMP, KET, and MER adsorbed C_{24} fullerene with solvent effect, respectively.

Table 4.4: Calculated value of Adsorption energy (E_{ad}), LUMO energy (E_{LUMO}), HOMO energy (E_{HOMO}), HOMO-LUMO gap (E_G), Recovery Time (τ) and Sensing Response (S) for AMP, KET, and MER adsorbed C_{24} fullerene with solvent effect, respectively.

System	E _{ad} (eV)	Elumo (eV)	Еномо (eV)	E _G (eV)	τ (sec)	S
C24	•••	-3.76	-5.59	1.83	••••	• • • • •
AMP/C ₂₄	-1.53	-2.63	-4.69	2.06	7.49×10^9	7310.04
KET/C ₂₄	-1.29	-2.56	-4.61	2.05	9.39×10^5	4964.69
MER/C ₂₄	-0.325	-3.74	-5.54	1.80	1.92× 10 ⁻¹⁰	2.19

Table 4.5: Comparison of the sensing property of C_{24} fullerene with other pristine nanomaterials.

AMP					
System	E _{ad} (eV)	d (ang)	Ref.		
C ₂₄ fullerene	-1.02	1.55	Our work		
BC ₃ nano-sheet	-0.46	1.70	27		
BC3 nano-tube	-0.68	1.66			
Al ₂₄ N ₂₄ nanocluster	-1.53		26		
Al ₁₂ N ₁₂ nano-cage	-1.01	2.38	22		
Al ₁₂ P ₁₂ nano-cage	-1.26	3.07			
Silicon carbide nanotube	-0.37	2.57	54		
(SiCNT)					
C ₆₀ Fullerene	-0.05	3.21	23		
	ME	ER			
System	E _{ad} (eV)	d (ang)	Ref.		
C ₂₄ fullerene	-0.365	2.95	Our Work		
B ₁₂ N ₁₂ nanoclusters	-0.62	2.36	31		
BN nanosheet	-0.13	3.87	30		
BN nanotube	-0.81	1.64			
BN nanocluster	-1.92	1.60			
B ₂₄ N ₂₄ nanocage	-0.67	1.65	29		
C ₇₀ fullerene	-0.26	3.1	28		
KET					
System	E _{ad} (eV)	d (ang)	Ref.		
C ₂₄ fullerene	-0.423	2.49	Our Work		
Functionalized single-walled	-4.16		32		
carbon nanotube-R1 Functionalized single-walled carbon nanotube-R2	-4.64				

4.3.1.8 Comparison of sensing property of C_{24} fullerene with other pristine nanomaterial

We compared the adsorption energies (E_{ad}) and interatomic distances (d) of illegal drug molecules AMP, KET, and MER over C_{24} fullerene with previously reported studies [22-32][54]to assess the versatility of pristine C_{24} fullerene in terms of its sensing mechanism

toward the drug molecules. **Table 4.5** makes it evident that the drug AMP has a relatively poor adsorption energy toward the pristine C_{60} fullerene with the wide interatomic distances, making it unsuitable for drug detection. A chemosorption with a significant interatomic distance is seen for the pristine $Al_{12}N_{12}$ and $Al_{12}P_{12}$ nano-cages, which restricts their utility for AMP drug detection. However, in contrast, our findings show that the C_{24} fullerene has a higher adsorption energy at the right interatomic lengths. Additionally, compared to functionalized single-walled carbon nanotube[32] and C_{70} fullerene,[28] KET and MER drug molecules had shorter interatomic distances and the best adsorption energy, suggesting that they are not suitable for drug detection purposes. Due to its physisorption towards these molecules and short recovery time, C_{24} fullerene may be a good candidate for sensing applications for KET and MER drug molecules, while AMP drug molecules may be removed.

4.3.2 Molecular Dynamics Simulation

The equilibrium and stability of drug molecules and C₂₄ fullerene complexes in the aqueous environment explicitly at room temperature (310K) are assured using molecular dynamics modeling, which is a very helpful tool. We noted from the DFT-PCM study previously described that the drug molecules' sensitivity to C₂₄ fullerene increases in the aqueous environment, which also improves C₂₄ fullerene's sensing response in comparison to the gaseous phase. Investigating such complexes' dynamical stability in a water environment is therefore required. Here, we have estimated the temperature, total energy, and root mean square deviation (RMSD) of the complexes up to 50 ns. **Figure 4.7** shows the

No. of Drug No. of C24 No. of water Dimension System Molecules fullerene molecule of the cubic box(nm) AMP/C₂₄ 4 2 4998 6.00×6.00 ×6.00 KET/C₂₄ 4 2 5677 6.00×6.00 ×6.00 MER/C₂₄ 4 2 4065 6.00×6.00 $\times 6.00$

Table 4.6: *Detail of the simulation boxes which used in this work*

snapshots from the MD simulation of AMP, KET, and MER drug molecules with C_{24} fullerene. The RMSD curves for the entire system against time are shown in **Figure 4.8(a)** in order to

determine the systems' initial equilibrium state. RMSD curves are shown for the three chosen systems AMP/C₂₄, KET/C₂₄, and MER/C₂₄. A close examination of the RMSD curves indicates

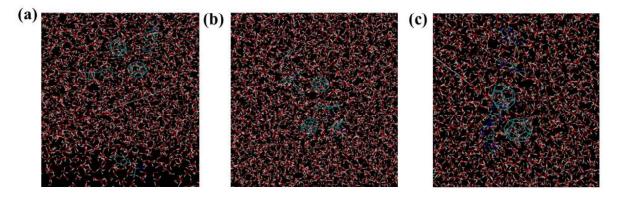


Figure 4.7: Captured moments in time for the simulation systems of (a) C_{24}/AMP , (b) C_{24}/KET , and (c) C_{24}/MER ..

that the all-equilibrated system saw very little variation throughout the 50 ns trajectory, which was about 4 nm. Comparing the AMP/ C_{24} system to KET/ C_{24} and MER/ C_{24} during the simulation, RMSD fluctuation for all three systems was kept to a minimum. To comprehend how drug molecules are distributed around C_{24} fullerene surfaces in water solvent for three systems, the radial distribution (RDF), or g(r), between drug molecules and C_{24} fullerenes has

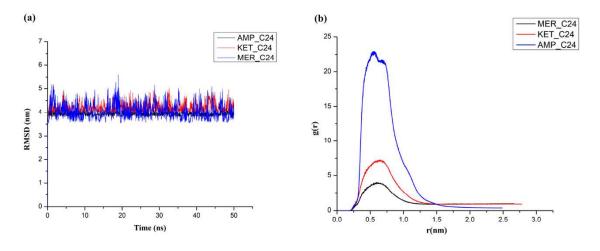


Figure 4.8: (a) Plots depicting the Root Mean Square Deviation (RMSD) over time for the drug molecules in interaction with C_{24} fullerene, and (b) a comparative analysis of the radial distribution (g(r)) of drug molecules in proximity to the surfaces of C_{24} fullerene, as a function of distance.

been determined (**Figure 4.8 (b)**). Notably, the center of gravity of drug molecules and C_{24} fullerenes have been plotted using the three figures. Based on the RDF graphs, we can infer that g(r) at short distances is zero due to the strong repulsive forces between drug molecules

and C₂₄ fullerenes. Remarkably, the larger size and heightened steric impact of the KET molecule resulted in the onset of nonzero values for g(r) occurring at a greater distance compared to the other two drug molecules, which also owe their delayed effect to their size and steric attributes. Due to the increased interaction energies of AMP and KET drug molecules with C₂₄ fullerene, which is compatible with DFT / PCM studies, the probability of finding the AMP and KET drug molecules surrounding the C₂₄ fullerene is higher than MER drug. The drug molecules are distributed over the surfaces of C₂₄ fullerenes in the range of 0.2–2.5 nm, as shown in Figure 8(b). The RDF results show that the adsorbed drug molecules and C₂₄ fullerene interact strongly at a distance of 0.5 nm for all three systems. In Figure 4.9(a) and (b), the fluctuation of total energy and temperature vs. simulation time steps of the drug/C₂₄ fullerene complexes has been plotted. During the simulation durations, we see that the total energies of three complexes approach the equilibrium state. The temperature variation graph shows that the simulated temperature of 310K was maintained for all complexes during the simulation. According to these results, the complexes approach equilibrium and adsorption for the drug/C₂₄ complexes occurs at room temperature during 50 ns. Fig.9(a) shows the maximum average total energy of the KET/C₂₄ and AMP/C₂₄ complexes -172193 and 152604 KJ/mol, respectively. Which are consistent with DFT/PCM results. In the water environment, KET/C₂₄ and AMP/C₂₄ showed greater stability.

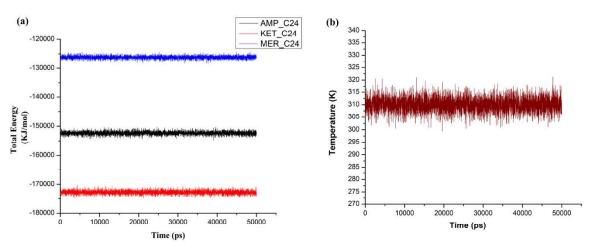


Figure 4.9: *MD* simulation plots for the (a) Total Energy and (b) Temperature for the drug molecules/ C_{24} against the time simulation.

4.4 Conclusions

Using the DFT-D3 method in conjunction with solvents, the present study investigated and analyzed the adsorption process of drug molecules on C₂₄ fullerene. According to our

findings, AMP and fullerene interact strongly in the gas phase, resulting in longer recovery times and stronger sensing responses. To put it differently, C_{24} fullerene can function as a chemical sensor to isolate AMP from a blend of various pharmacological compounds. Due to the physisorption nature of the adsorption of KET and MER on C_{24} fullerene, shorter recovery durations of 10^{-9} and 10^{-10} s were observed and an appreciable sensing response was observed. The interaction strengths and atomic separations between C_{24} and the drugs KET and MER are highly favorable, making C_{24} a suitable option for detecting these substances. Moreover, in aqueous environments, the AMP and KET complexes exhibit increased reactivity and stability, as their adsorption energy rises compared to gaseous phases, primarily attributable to the influence of the solvent. The pharmaceutical substances AMP and KET significantly accelerated the sensing response and recovery time. The removal of these medication compounds (AMP and KET) is thus possible using C_{24} fullerene in the water phase. As the solvent effect reduces the interaction energy of MER drug molecules, the MER/ C_{24} complex becomes less reactive and has a shorter recovery time, suggesting it can be used to detect MER.

In the water environment condition at room temperature (310K), the molecular dynamics simulation indicates that the complexes are well equilibrated and quite stable. A RDF plot also indicates that the drug molecules are well distributed across the C_{24} fullerene within acceptable distances, which supports the DFT results that indicate adsorption mechanisms.

As a result of the combination of DFT and MD simulation results, we concluded that C₂₄ fullerene would be an excellent choice for removing AMP and KET drug molecules in water environments and for detecting MER drug molecules at room temperature.

References:

- 1 J. Lenika and C. Wardak, *Procedia Eng.*, 2012, **47**, 144–147.
- 2 X. Li, P. Du, W. Zhang and L. Zhang, Curr. Opin. Environ. Sci. Heal., 2019, 9, 73–76.
- 3 F. MERZ, United Nations Off. Drugs Crime World Drug Rep. 2017, 2017, 85–86.
- 4 M. Carvalho, H. Carmo, V. M. Costa, J. P. Capela, H. Pontes, F. Remião, F. Carvalho and M. De Lourdes Bastos, *Arch. Toxicol.*, 2012, **86**, 1167–1231.
- 5 C. Sui, Q. Tu, N. Chen, B. Yan, Z. Lu, Z. Xu and Y. Zhang, Spectrosc. Lett., 2018, 51,

- 403-413.
- 6 R. Kohrs and M. E. Durieux, *Anesth. Analg.*, 1998, **87**, 1186–1193.
- 7 N. A. Anis, S. C. Berry, N. R. Burton and D. Lodge, *Br. J. Pharmacol.*, 1983, **79**, 565–575.
- 8 L. Wang and Z. Zhang, *Talanta*, 2008, **76**, 768–771.
- 9 M. Keyvanfard, V. Khosravi, H. Karimi-Maleh, K. Alizad and B. Rezaei, *J. Mol. Liq.*, 2013, **177**, 182–189.
- Z. Chen, G. Zhang, X. Chen, J. Chen, J. Liu and H. Yuan, *Biosens. Bioelectron.*, 2013, 41, 844–847.
- 11 R. Ahmadi, Int. J. Nano Dimens., 2018, 9, 325–335.
- 12 W. M. Mullett, *J. Biochem. Biophys. Methods*, 2007, **70**, 263–273.
- 13 R. D. Espy, S. F. Teunissen, N. E. Manicke, Y. Ren, Z. Ouyang, A. Van Asten and R. G. Cooks, *Anal. Chem.*, 2014, **86**, 7712–7718.
- T. Baciu, I. Botello, F. Borrull, M. Calull and C. Aguilar, *TrAC Trends Anal. Chem.*, 2015, **74**, 89–108.
- 15 M. L. Ochoa and P. B. Harrington, *Anal. Chem.*, 2004, **76**, 985–991.
- J. W. C. Alffenaar, E. M. Jongedijk, C. A. J. Van Winkel, M. Sariko, S. K. Heysell, S. Mpagama and D. J. Touw, *J. Antimicrob. Chemother.*, 2021, **76**, 423–429.
- 17 H. Dai, Y. Wang, X. Wu, L. Zhang and G. Chen, *Biosens. Bioelectron.*, 2009, **24**, 1230–1234.
- 18 H. H. Lin, Y. C. Li, C. H. Chang, C. Liu, A. L. Yu and C. H. Chen, *Anal. Chem.*, 2012, **84**, 113–120.
- 19 Y. Deng and G. Feng, *Anal. Chem.*, 2020, **92**, 14667–14675.
- 20 N. Ding, K. Liu, Y. Qi, C. Shang, X. Chang and Y. Fang, *Sensors Actuators, B Chem.*, 2021, **340**, 129964.
- 21 K. Vytřas, J. Pharm. Biomed. Anal., 1989, 7, 789–812.
- 22 H. R. A. El-Mageed and M. A. A. Ibrahim, J. Mol. Liq., 2021, 326, 115297.
- S. Bashiri, E. Vessally, A. Bekhradnia, A. Hosseinian and L. Edjlali, *Vacuum*, 2017, **136**, 156–162.
- 24 A. Qu and Y. Lin, *Phys. Lett. Sect. A Gen. At. Solid State Phys.*, 2020, **384**, 126726.
- Y. Yang, A. Sun and M. Eslami, *Phys. E Low-dimensional Syst. Nanostructures*, 2021, **125**, 114411.

- 26 M. M. R. Nayini, H. Sayadian, N. Razavipour and M. Rezazade, *Inorg. Chem. Commun.*, 2020, **121**, 108237.
- A. R. Moosavi-zare, M. Abdolmaleki, H. Goudarziafshar and H. Soleymanabadi, *Inorg. Chem. Commun.*, 2018, **91**, 95–101.
- 28 Y. Yang, A. Sun and W. Gu, Struct. Chem., 2021, 32, 457–468.
- Y. Yang and N. Ostadhosseini, *Phys. E Low-Dimensional Syst. Nanostructures*, 2021, 125, 114337.
- 30 S. A. Aslanzadeh, Mol. Phys., 2019, 117, 531–538.
- 31 H. R. Abd El-Mageed and H. S. Abbas, *J. Biomol. Struct. Dyn.*, 2021, **0**, 1–20.
- 32 R. Zhiani, I. Razavipanah and S. Emrani, *Struct. Chem.*, 2018, **29**, 1807–1815.
- 33 R. Singh and J. W. Lillard, Exp. Mol. Pathol., 2009, 86, 215–223.
- 34 S. Bibi, S. Ur-Rehman, L. Khalid, I. A. Bhatti, H. N. Bhatti, J. Iqbal, F. Q. Bai and H. X. Zhang, *RSC Adv.*, 2022, **12**, 2873–2887.
- 35 S. K. Jana, D. Chodvadiya, N. N. Som and P. K. Jha, *Diam. Relat. Mater.*, 2022, **129**, 109305.
- 36 M. J. Frisch, Trucks, G.W., Schlegel, H.B., Scuseria, G.E., Robb, M.A., Cheeseman, J.R., *Gaussian 09, Revis. A.02, Gaussian, Inc.*, 2016, 1–2.
- 37 S. Grimme, J. Antony, S. Ehrlich and H. Krieg, *J. Chem. Phys.*, 10.1063/1.3382344.
- 38 H. J. C. Berendsen, D. van der Spoel and R. van Drunen, *Comput. Phys. Commun.*, 1995, **91**, 43–56.
- 39 A. D. Mackerell, N. Banavali and N. Foloppe, *Biopolymers*, 2001, **56**, 257–265.
- 40 P. Mark and L. Nilsson, J. Phys. Chem. A, 2001, **105**, 9954–9960.
- 41 G. Bussi, D. Donadio and M. Parrinello, *J. Chem. Phys.*, 10.1063/1.2408420.
- 42 B. Hess, H. Bekker, H. J. C. Berendsen and J. G. E. M. Fraaije, *J. Comput. Chem.*, 1997, **18**, 1463–1472.
- 43 T. Darden, D. York and L. Pedersen, *J. Chem. Phys.*, 1993, **98**, 10089–10092.
- 44 K. S. William Humphrey, Andrew Dalke, *Journal Mol. Graph.*, 1996, **14**, 33–38.
- 45 C. A. Celaya, L. F. Hernández-Ayala, F. Buendía Zamudio, J. A. Vargas and M. Reina, *J. Mol. Liq.*,10.1016/j.molliq.2021.115528.
- 46 M. Rakib Hossain, M. Mehade Hasan, S. Ud Daula Shamim, T. Ferdous, M. Abul Hossain and F. Ahmed, *Comput. Theor. Chem.*, 2021, **1197**, 113156.

- 47 V. Nagarajan and R. Chandiramouli, *Mol. Phys.*, 10.1080/00268976.2021.1936248.
- 48 M. A. Hossain, M. R. Hossain, M. K. Hossain, J. I. Khandaker, F. Ahmed, T. Ferdous and M. A. Hossain, *Chem. Phys. Lett.*, 2020, **754**, 137701.
- 49 M. Shahabi and H. Raissi, J. Incl. Phenom. Macrocycl. Chem., 2015, 84, 99–114.
- 50 A. Hosseinian, E. Vessally, S. Yahyaei, L. Edjlali and A. Bekhradnia, *J. Clust. Sci.*, 2017, **28**, 2681–2692.
- 51 R. Chandiramouli, A. Srivastava and V. Nagarajan, *Appl. Surf. Sci.*, 2015, **351**, 662–672.
- 52 V. Nagarajan, R. Bhuvaneswari and R. Chandiramouli, *J. Mol. Graph. Model.*, 2023, **119**, 108395.
- 53 A. Ahmadi Peyghan, N. L. Hadipour and Z. Bagheri, *J. Phys. Chem. C*, 2013, **117**, 2427–2432.
- 54 Y. Yang, A. Sun and M. Eslami, *Phys. E Low-Dimensional Syst. Nanostructures*, 2021, **125**, 114411.
- P. Dipak, D. C. Tiwari, A. Samadhiya, N. Kumar, T. Biswajit, P. A. Singh and R. K. Tiwari, *J. Mater. Sci. Mater. Electron.*, 2020, **31**, 22512–22521.
- 56 H. Rahman, M. R. Hossain and T. Ferdous, J. Mol. Liq., 2020, **320**, 114427.
- 57 M. Cossi, V. Barone, R. Cammi and J. Tomasi, *Chem. Phys. Lett.*, 1996, **255**, 327–335.
- 58 M. Shahabi and H. Raissi, J. Incl. Phenom. Macrocycl. Chem., 2016, 86, 305–322.
- 59 J. Jiang, T. Yan, D. Cui, J. Wang, J. Shen, F. Guo and Y. Lin, *J. Mol. Liq.*, 2020, **315**, 113741.